Determination of blood indices of albino rats treated with aluminum chloride and investigation of antioxidant effects of vitamin E and C

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Abstract

The current study aims to investigate hematological and biochemical blood indices of albino rats administrated aluminum chloride (AlCl₃) for eight weeks, and to study the therapeutic effects of vitamin E and C. AlCl₃ decreased the total red blood cell count (by 18%), hemoglobin (7%) and hematocrit (20%), and increased white blood cell count (67%), lymphocytes (29%), mean corpuscular volume (14%), mean corpuscular hemaglobin (6%) and platelets (33%). Administration of vitamin E with or without vitamin C failed to restore levels of red blood cell counts, hematocrit, mean corpuscular volume, mean corpuscular hemaglobin or platelets, but vitamin E on its own restored levels of white blood cells, hemaglobin and lymphocytes.

AlCl₃ decreased serum glucose levels by 30%, and increased triglyceride (28%) and cholesterol (20%) levels; neither vitamin treatments restored the levels of these components. AlCl₃ increased levels of urea (12%), uric acid (77%) and creatinine (25%) compared to the controls, and vitamin E separately or together with vitamin C restored the levels of these nitrogen compounds.

The activities of alanine aminotransferase, alkaline phosphatase, and aspartate aminotransferase were also increased by the $AlCl_3$ treatment; the first two but not aspartate aminotransferase were restored by vitamin E separately or together with vitamin C.

We conclude that vitamin E separately or together with vitamin C suppressed cytogenetic injury and damage to some biochemical pathways of rat organs induced by AlCl₃.

Keywords: albino rats, aluminum chloride, blood indices, rat organs, vitamin E, vitamin C.

Introduction

Aluminum is a well-known toxic agent and represents a severe problem in a variety of medical (Nicolini *et al.* 1992) and environmental situations (Meranger 1989). The evidence implicating aluminium as a neurotoxin has been continuously mounting. Research on both animals and humans has linked it with neurocognitive dysfunction and in some cases death (Rifat *et al.* 1990). The major sources of aluminum include air, food and water (Michel 1990), and the gastrointestinal tract constitutes the main route of entry into the body. However, the absorption rate is low in normal human subjects (Brown *et al.* 1986).

Aluminum hydroxide, administered therapeutically in large quantities as an antacid and phosphate binder has been suggested to contribute to aluminum accumulation and toxicity (Lione 1985). Chronic exposition can cause alterations in skeletal, nervous, hematopoietic and respiratory systems (Chen *et al.* 2002; Cambell 2002). Blood urea is the principal end product of protein catabolism and a good indicator of kidney function. Uric acid is the end product of catabolism of purine bases; increased concentrations in the blood over the normal range might be due to extra degradation of purines in the liver, or an inability to excrete uric acid by the kidneys (Varely 1987). Creatinine appears in the serum in amounts proportional to the body's muscle mass and is more readily excreted by the kidneys than urea or uric acid (Pevicharova *et al.* 1997).

Blood enzymes are normally found in small amounts in circulation because of normal tissue turnover. Alanine aminotransferase as a liver enzyme significantly elevates in hepatobiliary disease, but also in connection with damage to the heart or skeletal muscle as well as liver parenchyma. Alkaline phosphatase is present on the cell surfaces in most human

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tissues, and belongs to a group of enzymes that catalyze the hydrolysis of phosphomonoesters at alkaline pH. High activity is found in the intestine, liver, bone, spleen and kidneys (Stryer 1995). Aluminum ions alter the properties and structure of cellular membranes, inhibiting many enzymes (Platt *et al.* 2001; Abreo & Glass 1993), and can act as antagonists for other elements such as calcium, magnesium, iron, silicon, phosphorus, copper and zinc (Ward *et al.* 2001).

Vitamin C is essential for the formation of collagen and intracellular material, bone, teeth and for the healing of wounds. It helps maintain elasticity of the skin aids the absorption of iron and improves resistance to infection. Vitamin E is the primary liposoluble antioxidant, perhaps important in scavenging free oxygen radicals and in stabilizing cell membranes, maintaining permeability (Packer 1993). Antioxidants such as vitamins E and C, coenzyme Q, glutathione and selenium ions can act synergistically, preventing lipid peroxidation and cell destruction (Escott-Stump & Mahan 2000).

The Gaza strip was exposed to Israeli bombing from Dec 27 2008 to Jan 18 2009, resulting in high concentrations of heavy metals. Such environmental contaminants can be transmitted to humans, causing many health complications (Manduca *et al.* 2009). Although many studies have been carried out on the toxic effect of aluminium ions and the antioxidant effects of vitamin E and C (Hayes *et al.* 2001; Eastmond *et al.* 2001; Manduca *et al.* 2009; Al-Faisal 2010), their effects on the body at a molecular level are still controversial. The present study investigates the different effects of AlCl₃ on blood indices of albino rats, and the subsequent response of rat tissues to therapeutic actions of vitamins E and C.

Materials & Methods

The study design involved one control and three treatment groups. It used 24 adult male albino rats, each weighing 100-120 gm, purchased from the breeding unit of the Biology Department, IUG. They were kept in plastic cages with wire mesh covers for one week before experimentation, and then divided in groups of six into one control and three treatment groups. Group one was administered 40 mg/l AlCl₃ dissolved in the drinking water (Fyiad 2007); group two was given 40 mg/l AlCl₃ plus vitamin E (150 mg/kg) (El-Nahas 1993); and group three had 40 mg/l AlCl₃ plus vitamins E and C (150 mg/kg) (El-Nahas 1993). Commercial balanced diet and water were continuously and regularly supplied *ad libitum* to the animals throughout the experimental period. The duration of the experiment was 8 weeks, when blood samples were collected from the jugular vein for hematological and biochemical examination.

Routine hematological parameters and a complete blood count was carried out using an automated 18-parameter hematology analyzer (ABX Micros 60, Horiba ABX, France). Clear serum samples were separated by centrifugation at 3000 rpm. for 20 min, collected and stored in a deep freeze at (-20 0 C) for biochemical analysis. Glucose, triglyceride and cholesterol were determined using classical methods described by Trinder (1979), Fossati & Prencipe (1982) and Allain *et al.* (1974), respectively. Serum urea measurement was based on cleavage of urea with urease (Fawcett & Scott 1960). Serum uric acid was determined according to Fossatti *et al.* (1972). Activities of serum aspartate aminotransferase and alanine aminotransferase were determined according to the classic method of Reitman & Frankel (1957); measurement of serum alkaline phosphatase activity was also based on the method of Bessey *et al.* (1946).

Data were analyzed using SPSS version 13 for Windows. ANOVA was used to test for differences among groups; differences were considered significant if p < 0.05.

Results

Table 1 summarizes the effect of $AlCl_3$ and vitamins C and E on hematological parameters. After eight week of $AlCl_3$ administration, there was significant decrease in the total red blood cells, red blood cells, hemoglobin and hematocrit compared to the control. In contrast, white blood cells, lymphocytes, corpuscular volume, corpuscular hemoglobin and platelets showed a significant increase compared to the control. There was a non-significant increase in corpuscular hemoglobin concentration. Administration of vitamin E alone, or with vitamin C, failed to counteract the effect of $AlCl_3$ on red blood cells, hematocrit, corpuscular volume, corpuscular hemoglobin, corpuscular hemoglobin concentration and platelets. Vitamin E alone counteracted the effect of the ion on white blood cells, lymphocytes and hemoglobin.

Parameter	control	AlCl ₃	AlCl ₃ + vitamin E	AlCl ₃ + vitamins E & C
White blood cells (x 10^3 cell/µl)	$3.90^b\pm0.19$	$6.5^{a}\pm0.24$	$4.10^b\pm0.36$	$5.85^a\pm0.29$
Lymphocytes	$60.6^{b}\pm2.5$	$78.2^{a}\pm3.0$	$62.6^b\pm3.2$	$75.9^{a}\pm2.3$
Red blood cells (x 10^6 cell/µl)	$11.19^{a}\pm0.20$	$9.14^{b}\pm0.35$	$9.25^{\text{b}} \pm 0.30$	$9.51^{\text{b}}\pm0.31$
Hemoglobin (g/dl)	$16.26^{a}\pm0.70$	$15.18^{b}\pm0.18$	$16.18^{a}\pm0.19$	$15.45^b\pm0.20$
Hematocrit (%)	$65.9^{\rm a}\pm1.1$	$53.0^{\text{b}} \pm 1.2$	$52.5^{\text{b}} \pm 1.3$	$51.6^{\text{b}} \pm 1.3$
Corpuscular volume (fi)	$14.5^{c}\pm0.2$	$16.6^{b}\pm0.2$	$17.4^{b}\pm0.3$	$16.2^{b}\pm0.2$
Corpuscular hemoglobin (pg)	$27.14^{b}\pm0.20$	$28.65^a\pm0.16$	$30.84^{a}\pm0.19$	$29.97^{a}\pm0.23$
Corpuscular hemoglobin concentration (g/dl)	$53.52^b\pm0.25$	$57.95^{a}\pm0.31$	$56.70^a\pm0.36$	$54.20^{a}\pm0.42$
Platelets (x $10^3/\mu l$)	$595.1^{c}\pm23.3$	$790.0^{a}\pm25.2$	$731.8^{b}\pm31.3$	$723.0^b\pm33.9$

Table 1: Hematological indices of the rats administrated $AlCl_3$, vitamin E and vitamin C (all values expressed as mean \pm SE). Means with different subscripts in the same row differ significantly (p<0.05).

Table 2 shows changes in glucose, triglycerides and cholesterol concentrations in the experimental groups. Aluminium treatment significantly decreased serum glucose levels (by 30%) and increased significantly triglycerides (28%) and cholesterol (20%). Treatments with vitamin E alone or with vitamin C did not restore these compounds to control levels.

Parameter	control	AlCl ₃	AlCl ₃ + vitamin E	AlCl ₃ + vitamins
				E & C
Glucose (mg/dl)	$95.2^{a} \pm 3.1$	$66.8^{b} \pm 2.2$	$68.8^{b} \pm 2.2$	$67.5^{b} \pm 2.3$
Triglycerides(mg/dl)	$88.5^{b} \pm 2.5$	$113.3^{a} \pm 2.3$	$105.7^{a} \pm 3.4$	$103.7^a\pm2.2$
Cholesterol (mg/dl)	$129.1^{\circ} \pm 2.3$	$155.0^{a} \pm 3.2$	$152.2^{a} \pm 3.3$	$143.6^{b} \pm 2.2$
Urea (mg/dl)	$25.3^{b} \pm 0.1$	$28.3^{a} \pm 1.1$	$26.5^{b} \pm 1.2$	$25.4^{b} \pm 2.1$
Uric acid (mg/dl)	$3.51^{b} \pm 0.20$	$6.21^{a} \pm 0.26$	$3.77^{b} \pm 0.24$	$4.11^{b} \pm 0.25$
Creatinine (mg/dl)	$0.60^{ m b} \pm 0.01$	$0.75^{\mathrm{a}} \pm 0.02$	$0.65^{b} \pm 0.01$	$0.62^{b} \pm 0.02$
Alanine aminotransferase (IU/ml)	$25.5^{b} \pm 0.3$	$30.8^{a} \pm 0.4$	$28.1^{b} \pm 0.3$	$27.4^{\rm b} \pm 0.3$
Aspartate amino transferase (IU/ml)	$21.9^{\circ} \pm 0.4$	$31.4^{a} \pm 0.4$	$28.7^{b} \pm 0.5$	$29.7^{\mathrm{b}}\pm0.6$
alkaline phosphatase (IU/ml)	$81.5^{b} \pm 0.6$	$100.1^{a} \pm 4.3$	$90.1^{b} \pm 2.2$	$88.2^{b} \pm 2.2$

Table 2: Chemical concentrations in albino rats administrated $AlCl_3$, vitamin E and vitamin C
(all values expressed as mean \pm SE). Means with different subscripts in the same row
differ significantly (p<0.05).</th>

Levels of non-protein nitrogenous constituents for treatment groups are also given in Table 2. The aluminium treatment significantly increased urea (by 12%), uric acid (77%) and creatinine

levels (25%) compared to the control. Vitamin E separately or together with vitamin C significantly counteracted the effects of aluminium.

Activities of serum aspartate amino transferase, alanine aminotransferase and alkaline phosphatase (Table 2) increased significantly following aluminium treatment. As for the nitrogenous compounds, these increases were counteracted by treatment with vitamin E alone or together with vitamin C. In contrast, aspartate amino transferase activity was not counteracted at all.

Discussion

This study aimed to determine the toxic effects of the aluminium ion, and the therapeutic effects of vitamin E and vitamin C, on rats. We found highly significant decreases in hemoglobin, red blood cells and hematocrit among aluminium-treated rats, as have others (Karmaker *et al.* 2000). The reduction in hemoglobin content might be due to increased rate of destruction or reduction in the rate of formation of red blood cells. This intepretation was supported by the low levels of red blood cells in the treated groups. Reductions in hematocrit, red blood cells and hemoglobin might be attributed to hyperactivity of bone marrow, leading to production of red blood cells with impaired integrity that are easily destroyed in the circulation (Karmaker *et al.* 2000). On the other hand, these decreases could alternatively reflect a lower oxygen supply to different tissues, resulting in low energy production in the rats. The decrease in hemoglobin could be not only due to decrease in red blood cells count but also to impaired biosynthesis of heme in the bone marrow.

The significant increase in white blood cell levels of aluminium-treated rats might indicate activation of the immune system, a normal cell-mediated immune response (El-Demerdash 2004). The increase in lymphocytes could be due to the toxic action of the aluminium ion that stimulates the hemopoietic system to release more of these cells, causing an increase in their number in the blood stream.

The increase in corpuscular volume, corpuscular hemoglobin, corpuscular hemoglobin concentration and platelets were consistent with changes in red blood cell counts and hemoglobin levels. These changes may be correlated with some pathological changes developed in blood-forming organs, or with the destruction of red blood cells, or with both factors. In this regard, from similar results Naylor (1971) concluded that anemia resulted from hemodilation, extra vascular hemolysis and toxic dyshemopiosis.

Our findings show that vitamin E on its own counteracted the effects of the aluminium ion on white blood cells, lymphocytes and hemoglobin. Vitamin E and vitamin C separately increase the activities of antioxidant enzymes in various tissues of rats, especially liver tissues and also vitamin E on bone marrow, where the different blood cells are formed (Shireen *et al.* 2008).

Our findings also revealed a decrease in serum glucose levels in response to aluminium. Indirectly, aluminium is known to play a specific role in carbohydrate metabolism (Thirunavukkarasu & Sakthisekaran 2003). Concerning lipid metabolism, our results demonstrated that triglycerides and total cholesterol levels increased in response to aluminium, consistent with increasing lipogenesis in the liver (Thirunavukkarasu & Sakthisekaran, 2003). Vitamin E separately or together with vitamin C could not counteract the effects of the aluminium ion on glucose, cholesterol and triglycerides. This indicates that vitamin E is only indirectly involved in metabolism of these compounds via defending the integrity of cells against oxidating agents. Vitamin E mediates mitochondrial superoxide generation, suggesting a possible mode of action at tissue level, and it also modulates the expression and/or activation of redox-sensitive biological response modifiers that regulate important cellular events (Chow 2004).

Enhanced protein catabolism and accelerated amino acid deamination in response to low glucose levels caused by aluminium ion administration is the best interpretation for the elevated levels of urea. The presence of toxic compounds can increase blood urea and decrease plasma protein (Berne & Levy 1998). The observed increase in uric acid concentration might be due to extra degradation of purines in the liver, or an inability to excrete uric acid by the kidneys (Varely 1987). An increase in creatinine has been seen, interpreted as caused by a decrease in muscle mass (Pevicharova *et al.* 1997) or abnormal glomerular function of the kidneys induced by AlCl₃ administration (Berne & Levy 1998). The observation that vitamin E separately or together with vitamin C counteracted the toxic effects of the aluminium ion in forming these nitrogenous compounds indicates that the vitamins reverse and thus inhibit interactions with metabolic enzymes involved in their synthesis in the liver and muscles.

Serum transaminases (aspartate amino transferase and alanine aminotransferase) and alkaline phosphatase exhibited a significant increase in treated rats, perhaps indicating persistent cellular injury (Bansal *et al.* 2005). Elevated activities of serum transaminases could be a sign of impaired liver function. Alkaline phosphatase has a specific location within both sinusoidal and bile canalicular membranes, accounting for its more predominant elevation in certain disorders (Bansal *et al.* 2005): acute cell necrosis liberates alkaline phosphatase into the blood circulation and its level is elevated. As with biosynthetic enzymes of nitrogen compounds, vitamin E separately or together with vitamin C reversed the toxic effects of aluminium ions on the activities of alkaline phosphatase and alanine aminotransferase, but not aspartate amino transferase. Their effect on aspartate amino transferase requires further investigation.

We conclude that aluminum ions significantly decrease red blood cell counts, hemoglobin, hematocrit and glucose, and significantly increase white blood cell counts, lymphocytes, corpuscular volume, corpuscular hemoglobin, corpuscular hemoglobin concentration, platelets, triglycerides, cholesterol, urea, uric acid, creatinine and the activity levels of alanine aminotransferase, aspartate amino transferase and alkaline phosphatase. Vitamin E on its own counteracts the effect of AlCl₃ on white blood cell counts, hemoglobin and lymphocytes. Vitamin E alone or together with vitamin C counteracts the effects of aluminium ions on urea, uric acid, creatinine and on the activities of alanine aminotransferase and alkaline phosphatase. Consequently, vitamin E separately or together with vitamin C suppresses cytogenetic injuries and damage to some biochemical organ pathways (e.g. liver, kidney, bone marrow) induced by AlCl₃ administration.

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References

- Abreo K & Glass J (1993) Cellular, biochemical and molecular mechanisms of aluminium toxicity. Nephrology Dialysis Transplantation 8(1): 5
- Al-Faisal AHM, Hussein AM & Abdul Kaleg AR (2010) Estimation of DNA damage, cytotoxicity and antioxidant status of heavy metals and benzene among petrol workers in Baghdad-Irag. Isan Journal of Pharmaceutical Sciences 6(1): 85-94
- Allain CC, Poon LS & Chan CSG (1974) Enzymatic determination of total serum cholesterol. Clinical Chemistry 20 (4): 470-75
- Bansal AK, Bansal M & Soni G (2005) Protective role of Vitamin E pre-treatment on N-nitrosodiethylamine induced oxidative stress in rat liver. Chemico-Biological Interactions 156: 101-111
- Bartels H, Bohmer M & Heierli C (1972) Serum creatinine determination without protein precipitation. Clinica Chemica Acta 37: 193-197
- Berne MR & Levy NM (1998) Physiology. 4th ed. Mosby, New York. pp 910-929
- Bessey OA, Lowry DH & Brock JM (1946) Method for the determination of alkaline phosphatase with five cubic milliliters of serum. Journal of Biological Chemistry 146: 321

- Brown S, Mendoza N & Bertholt RK (1986) Absorption of aluminum from aceglutamide in healthy adult males. Research Communications in Chemical Pathology & Pharmacology 53:105–116
- Cambell A (2002) The potential role of aluminum in Alzheimer's disease. Nephrology Dialysis Transplantation 17: 2-17
- Chen J, Wang M & Run D (2002) Early chronic Al exposure impairs long-term potentiality and depression to the rat dentate gurus in vivo. Neuroscience 112 (4): 879
- Chow CK (2004) Biological functions and metabolic fate of vitamin E. Journal of Biomedical Science 11: 295-302
- Eastmond DA, Schuler M & Franz CH (2001) Characterization and mechanisms of chromosomal aberrations induced by benzene in mice and humans. Research Reports of the Health Effects Institute 103: 69-80
- El-Demerdash FM (2004) Antioxidant effect of vitamin E and selenium on lipid peroxidation, enzyme activities and biochemical parameters in rats exposed to aluminium. J. Trace Elements in Medical Biology 18: 113-122
- El-Nahas SM, Mattar FE & Mohamed AA (1993) Protective effects of vitamin C and E. Mutation Research 301:143
- Escott-Stump S & Mahan LK (2000) Krause's Food, Nutrition, and Diet Therapy. 10th ed. WB Saunders, Philadelphia, PA
- Fawcett JK & Scott JE (1960) A rapid and precise method for the determination of urea. Journal of Clinical Pathology 13:156-159
- Fossati P & Prencipe L (1982) Serum triglycerides determined colorimetrically with an enzyme that produces hydrogen peroxide. Clinical Chemistry 28 (10): 2077-80
- Fossatti P, Prencipe L & Berti G (1980) Use of 3,5-dichloro-2-hydroxy-benzenesulfonic acid/4-aminophenazone chromogenic system in direct enzymic assay of uric acid in serum and urine. Clinical Chemistry 26: 227-31
- Fyiad AA (2007) Aluminum toxicity and oxidative damage reduced ion by melatonin in rats. Journal of Applied Scientific Research 3(10): 1210-1217
- Hayes RB, Songnian Y, Dosemeci M & Linet M (2001) Benzene and lymphohematopoietic malignancies in humans. American Journal of Industrial Medicine 40(2): 117-26
- Karmakar R, Bhattacharya R & Chatterjee M (2000) Biochemical, haematological and histopathological study in relation to time-related cadmium-induced hepatotoxicity in mice. Biometals 13(3): 231-239
- Lione A (1985) Aluminum toxicology and the aluminium-containing medications. Pharmacology &Therapeutics 29: 255–285
- Manduca P, Barbieri M & Barbieri M (2010) Analysis of pilot survey of metal content in samples of hair collected in December 2009 in Gaza. http://www.newweapons.org.,.
- Meranger JC (1989) How aluminium levels in subsurface drinking water supplies in Canada can be used to predict possible impact by acidic deposition. pp. 107–116 in Lewis, T.E.(ed) *Environmental Chemistry and Toxicology of Aluminium*. Lewis Publications, Chelsea, Ml.
- Michel P, Commenges D & Dartigues JF (1990) Study of the relationship between Alzheimer's disease and aluminum in drinking water. Neurobiology of Aging 11:264
- Naylor S (1971) The hematology and histopathology of *Trypanosoma congolense* infection in cattle. Tropical Animal Health & Production 3: 159–168
- Nicolini M, Zatta PF & Corain B (1992) Aluminum in Chemistry, Biology and Medicine. Cortina Intern, Verona and Raven Press, New York.
- Packer L (1993) Vitamin E: biological activity and health benefits: Overview. p. 977-982 in Packer L & Fuchs J (eds.) *Vitamin E in health and disease*. New York, Marcel Dekker, Inc.
- Pevicharova GT, Dimova PI & Atanasova-Goranova VK (1997) Effect of food products on endogenous generation of nitrosamines in rats. British Journal of Nutrition 78(2):325-45
- Platt B, Fiddler G, Riedel G & Henderson Z (2001) Al toxicity in the rat brain: histochemical and immunocytochemical evidence. Brain Research Bulletin 55 (2), 257
- Reitman S & Frankel S (1957) A colorimetric method for the glutamic-pyruvate transaminase. American Journal of Clinical Pathology 28: 56-63
- Rifat SL, Eastwood MR & Crapper McLachlan DR (1990) Effects of exposure of miners to aluminum powder. Lancet 336:1162–1165
- Shireen KF, Pace RD & Mahboob M (2008) Effects of dietary vitamin E, C and soybean oil supplementation on antioxidant enzyme activities in liver and muscles of rats. Food & Chemical Toxicology 46: 3290-4
- Stryer L (1995) Biochemistry. 4th ed. W.H. Freeman and Company, New York, USA. pp. 607-610
- Thirunavukkarasu C & Sakthisekaran D (2003) Influence of sodium selenite on glycoprotein contents in normal and N-nitrosodiethylamine initiated and phenobarbital promoted rat liver tumors. Pharmacological Research 48:167-173
- Trinder P (1969) Glucose GOD-PAP method Enzymatic colorimetric method. Annals of Clinical Biochemistry 6: 24-27

Varely H (1987) *Practical clinical biochemistry*. 6th ed. Gowenlock AH, McMurray JR & McLauchlan DM (eds). London, Heinemann Medical Books. pp 477-549

Ward RJ, Zhang Y & Crichton RR (2001) Aluminium toxicity and iron homeostasis. Journal of Inorganic Biochemistry 87(1-2): 9

الملخص العربي

تحديد المؤشرات الخاصة بدم الفئران البيضاء المعالجة بكلوريد الألومنيوم للتحقق من الآثار المضادة للأكسدة لفيتاميني (ج) و (هـ)

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الملخص العربى

تهدف الدراسة الحالية إلى التحقق من المؤشرات الدموية والكيميائية الحيوية لدم الفئران البيضاء المعاالجة بكلوريد الألومنيوم (AlCl3) لمدة ثمانية أسابيع لدراسة الآثار العلاجية لفيتاميني (ج) و (هـ). يؤدي كلوريد الألومنيوم إلى انخفاض عدد خلايا الدم الحمراء (بنسبة 18 ٪) والهيموجلوبين (بنسبة 7 ٪) والهيماتوكريت (بنسبة 20 ٪) ، كما يؤدي لزيادة عدد خلايا الدم البيضاء (بنسبة 67 ٪) والميموجلوبين (بنسبة 7 ٪) ومتوسط حجم خلايا الدم (بنسبة 10 ٪) ومتوسط المحتوى الهيموجلوبيني لخلايا الدم (بنسبة 6 ٪) أو الصفائح الدموية (بنسبة 30 ٪) . فشل فيتامين (هـ) عدد إلى المحتوى المحتوى الهيموجلوبيني لخلايا الدم (بنسبة 6 ٪) أو الصفائح الدموية (بنسبة 33 ٪) . فشل فيتامين (هـ) عند إعطائه الفئران وحده أو مع فيتامين (ج) في استعادة المستويات الطبيعية لأعداد خلايا الدم الحمراء والهيماتوكريت ومتوسط حجم الخلايا وكذا متوسط المحتوى الهيموجلوبيني لخلايا الدم أو الصفائح الدموية (بنسبة 33 ٪) . فشل فيتامين (هـ)

عمل كلوريد الألمونيوم على خفض مستويات السكر في الدم بنسبة 30 ٪ وزيادة مستويات الدهون الثلاثية (بنسبة 28 ٪) والكولسترول (بنسبة 20 ٪) ولم يسهم العلاج بأياً من الفيتامينين في استعادة مستويات تلك المكونات، كما أسهم في زيادة مستويات اليوريا (بنسبة 12 ٪) وحمض اليوريك (بنسبة 77 ٪) والكرياتينين (بنسبة 25 ٪) للفئر ان المعالجة به مقارنة بالفئر ان غير المعالجة، إلا أن فيتامين (هـ) على حدة أو بالاشتراك مع فيتامين (ج) قام باستعادة المستويات الطبيعية لهذه المركبات النيتروجينية.

من الملحوظ أن نشاط إنزيمات الألانين أمينوتر انسفيريز والألكالاين فوسفاتيز والأسبارتيت أمينوتر انسفيريز قد زاد نتيجة للمعالجة بكلوريد الألومنيوم، إلا أن أول إنزيمين (ما عدا الأسبارتيت أمينوتر انسفيريز) قد تأثرا بالعلاج بفيتامين (هـ) وحده أو مع فيتامين (ج) مما أدى إلى إستعادة النشاط الطبيعي.

مما سبق فإننا نُخلص إلى أن فيتامين (هـ) يمكنه بشكل منفصل أو مع فيتامين (ج) وقف الإصابات الخلوية والأضرار التي تلحق ببعض المسارات الحيوية لأعضاء الفئران والتي تتسبب فيها مادة كلوريد الألمونيوم.