

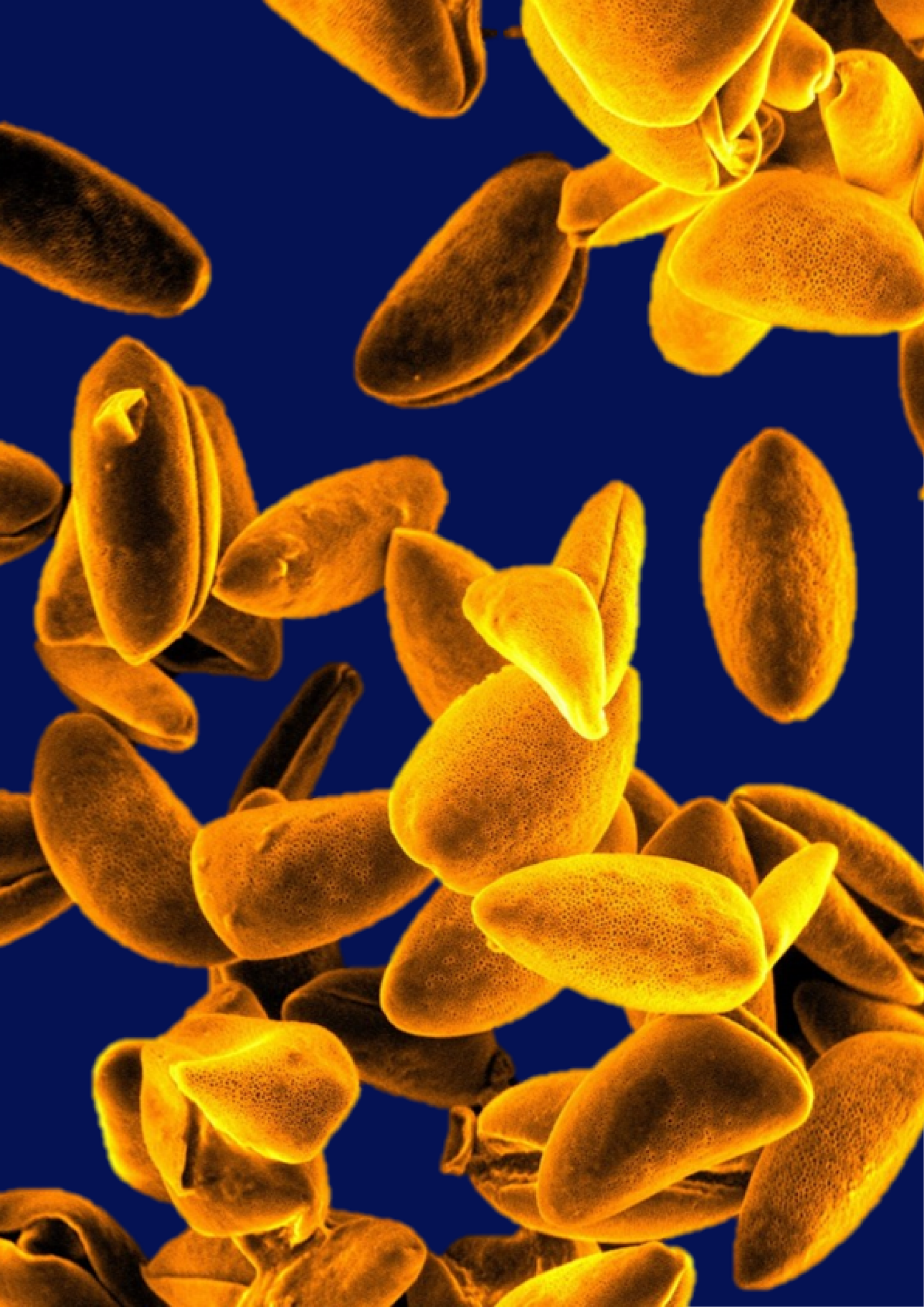


University of
Nottingham

Nanoscale and Microscale Research Centre

nmRC Facilities





About the nmRC

We are an inter-disciplinary facility dedicated to supporting and promoting world-leading nanoscience and materials characterisation.

The nmRC is a hub for state-of-the-art facilities and the allied expertise needed for materials imaging, chemical imaging, compositional analysis and nanofabrication.

The Centre is proud to support research and analytical services from across the engineering, physical and life sciences for all at the University of Nottingham and beyond.



Facilities in this brochure:

- Scanning electron microscopy (SEM)
- In-situ SEM techniques
- Transmission electron microscopy (TEM)
- Raman spectroscopy
- X-ray photoelectron spectroscopy (XPS)
- Secondary ion mass spectrometry (ToF-SIMS & 3D OrbiSIMS)
- Confocal microscopy
- Cryogenic materials characterisation
- Atomic force microscopy (AFM)
- Nanofabrication suite
- Biophysical analysis
- Particle sizing suite
- Liquid chromatography-mass spectrometry (LC-MS)



Scanning electron microscopy (SEM)

Scanning electron microscopy uses a beam of electrons to characterise the topography and structure of a sample. High depth of field images with a 3D perspective and lateral resolutions down to 1-nm can be supplemented with simultaneous chemical and mechanical data.

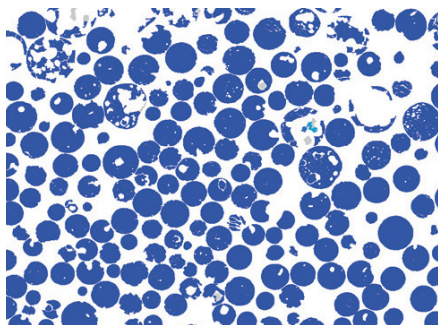
Variations of SEM allow for analysis options and sample manipulations that can accommodate a variety of sample types and formats.

Capabilities

- High magnification imaging of conducting and non-conducting materials
- Microscale topography and morphology analysis
- Elemental and compositional analysis
- Surface feature identification and measurement
- Sample sectioning, cryo-handling and micro-manipulation (FIB-SEM)
- Crystallographic analysis, micro-electron diffraction
- Wet, uncoated or dynamic environmental analysis (ESEM)

Typical applications

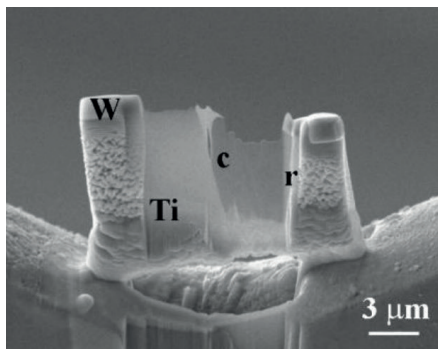
- Manufacturing analytics (stress and strain artefacts, wear and failure analysis and others)
- Micro- and nano-structural imaging and component identification
- Sample sectioning and high magnification full-structure visualisation
- Micro- and nano-particle/system sizing and localisation
- Grain sizing and qualitative/quantitative elemental mapping



Mineral mapping of compositionally uniform, magnetic microspheres

Mineral liberation analysis (MLA) enables automated large area analysis of sectioned samples in order to identify and quantify mineral distribution and composition. Image adjacent reveals high homogeneity for dense and porous, magnetic microspheres. A total of 1501 particles (99.7 wt%) were quantified and classified as $\text{Ca}_2\text{Fe}_2\text{O}_5$. Notably, this technique can be used to obtain a statistically significant set of data.

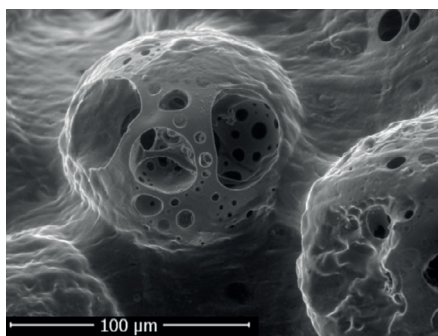
J Molinar Díaz, SA Samad, E Steer, N Neate, H Constantin, MT Islam, PD Brown, I Ahmed. *Materials Advances* (2020), DOI: 10.1039/D0MA000564A.



FIB sectioning of a biological Cryo-SEM sample

FIB-SEM manipulation allows delicate sample processing, such as cross sectional milling and 'lift-outs' which can in turn be imaged, or removed for analysis with further techniques (such as Raman spectroscopy, ToF-SIMS and TEM). This figure shows a lift-out section of a biomaterial interface between a Human Osteoblast (HOb) cell and a Titanium (Ti) foil. The lift-out section from a cryogenically frozen sample can be seen to contain a cell (c), resin (r), Ti and Tungsten (W) and is attached to a TEM support grid ready for further higher resolution analysis.

Microscopy at the life sciences/physical sciences interface. Paul D Brown, Hannah K Edwards and Mike W Fay. *Journal of Physics: Conference Series* 241 (2010), 012019



ESEM of phosphate-based porous microspheres interacting with mesenchymal cells

Environmental SEM (ESEM) was utilised to image phosphate-based glasses, porous microspheres (with Mg and Ti modifications, in this case M24T0) interacting with human mesenchymal cells. The technique allows the investigation of these materials in a hydrated or uncoated state. This is necessary to visualise the cellular interactions of the microspheres to assess growth and osteogenic potential. The image to the right shows cells migrating inside the pores of the microspheres in cell cultures on day 21.

MT Islam, L Macri-Pellizzeri, KMZ Hossain, V Sottile, I Ahmed. *Materials Science and Engineering: C* 120 (2021), 111668.

Our facilities

Zeiss Crossbeam 550 (HR-CAT-SEM): The high resolution cryogenic analytical and transfer (HR-CAT-SEM) instrument is a cryo-FIBSEM system with Field Emission Gun (FEG) optics that enables the highest levels of spatial resolution with cutting edge in-situ preservation and processing of a large variety of materials including biological, wet and magnetic samples. Extensive peripherals including a gas injection system (GIS), STEM detector and electron backscatter detector (EBSD) enable holistic interrogation of materials.

JEOL 7100F FEG-SEM: FEG-SEM for nanometre resolution imaging, with energy dispersive X-ray spectroscopy (EDS), wavelength dispersive X-ray spectroscopy (WDS) and electron backscatter diffraction (EBSD) capabilities in addition to a heating stage for exhaustive characterisation.

ThermoFisher (FEI) Quanta200 3D DualBeam FIB/SEM: FIB-SEM with cryogenic capability especially suited to biological samples and complex sample manipulations or lift out preparation.

ThermoFisher (FEI) Quanta 650 ESEM: State of the art ESEM for the imaging of uncoated or wet samples in an air, water vapour or nitrogen environment with a high sensitivity EDS detector. A Peltier cooling stage allows for dynamic analysis with temperature and relative humidity control.

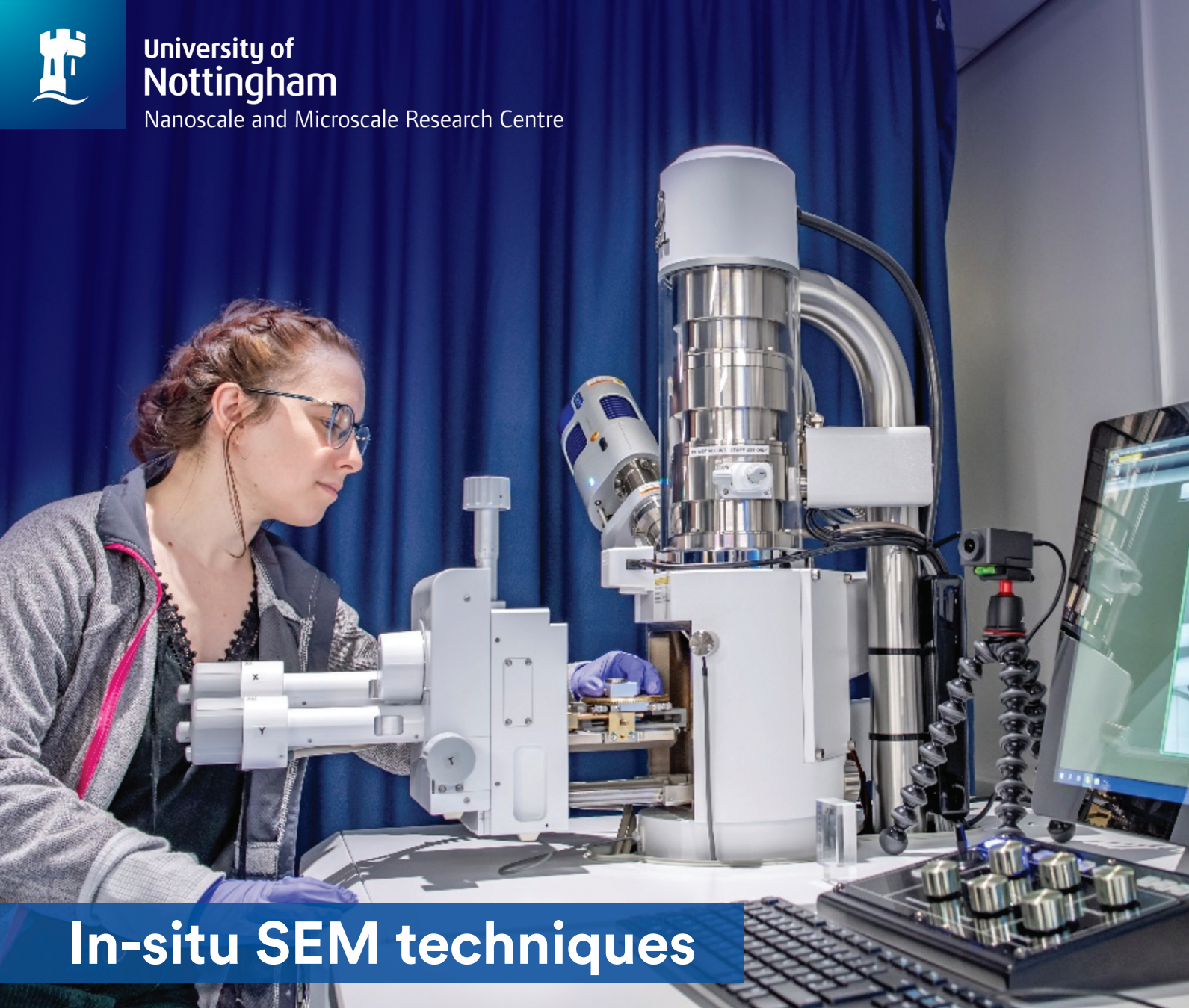
ThermoFisher (FEI) Quanta 600: Performs fully automated, large area and high-resolution analyses of polished sample specimens. Used to identify and quantify mineral composition and distribution (Mineral Liberation Analysis) with the provision of complex statistical analytics.

Also available... JEOL 7000F, ThermoFisher (FEI) XL30 SEM, JEOL JSM IT-200 SEM, JEOL 6490LV SEM.

Find out how SEM could help with your applications, designs or solutions:

nmrcenquiries@nottingham.ac.uk | +44 (0)115 951 5046

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In-situ SEM techniques

In-situ techniques allow for the examination and manipulation of samples within the scanning electron microscope (SEM), providing real-time insights into material properties and behaviours. With capabilities ranging from environmental imaging and elemental analysis to crystallographic studies and focussed ion beam processing, they are essential tools for detailed and dynamic investigation of materials in their native states.

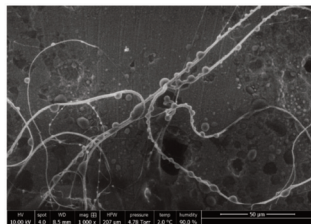
Capabilities

- Environmental SEM (ESEM)
- Backscattered electron imaging (BSE)
- Energy-Dispersive X-ray Spectroscopy (EDS)
- Wavelength-Dispersive X-ray Spectroscopy (WDS)
- Mineral Liberation Analysis (MLA)
- In-situ stages for heating, microtensile testing, and nanoindentation
- Focussed Ion Beam Scanning Electron Microscopy (FIB-SEM)
- Electron Backscatter Diffraction (EBSD)
- Scanning Transmission Electron Microscopy (STEM) in a SEM

Techniques:

Environmental SEM

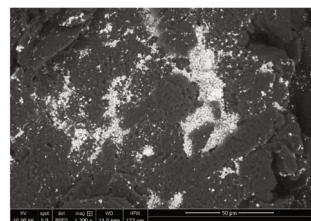
ESEM allows examination of fully hydrated 'wet' samples and of poorly conductive uncoated materials, none of which can be imaged in the high vacuum conditions of a conventional SEM. Pressure and temperature can be varied within the chamber, and a range of gases can be used (such as water vapour or nitrogen), to simulate experimental conditions.



Microdroplets of water formed on spider webbing via ESEM control of pressure (4.78 Torr) and temperature (2 °C) to form humidity (90%) within the chamber. Data courtesy of Prof. Sara Goodacre (School of Life Sciences) and Ms Nicola Weston.

Backscattered Electron imaging

BSE imaging is complementary to EDS/WDS showing atomic number (Z) contrast and highlighting differences in the composition of different phases based on the BSE response.

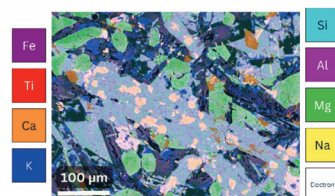


BSE image of copper oxide (CuO) within a darker polymer matrix highlighting major Z contrast differences, confirmed by EDS.

Data courtesy of Ms Nicola Weston.

Energy-Dispersive X-ray Spectroscopy

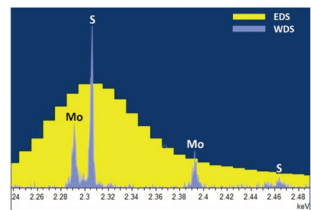
EDS measures the energy of X-rays emitted from elements in a material, and is used to analyse the elemental composition of them within the electron microscope. If materials are suitably prepared, it is also possible to obtain quantitative elemental analysis.



EDS map of a Basaltic Dyke intrusion from the Isle of Skye. Data courtesy of Ms Lorelei Robertson.

Wavelength-Dispersive X-ray Spectroscopy (WDS)

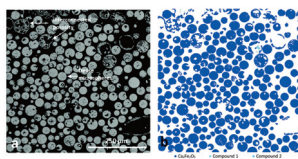
WDS differs from EDS as it measures the wavelengths of emitted X-rays producing several benefits: lower detection limits and therefore better for trace/minor elements (<0.5wt%), light element detection, and much improved spectral resolution minimising peak overlaps typically evident in EDS allowing the collection of "cleaner" X-ray maps.



EDS (yellow) and WDS (lilac) overlay of a molybdenum sulphide nanomaterial. The WDS spectrum resolves the Mo and S peaks that overlap in the EDS spectrum. Data courtesy of Dr Luke Norman

Mineral Liberation Analysis

MLA utilises two EDS detectors to allow automated large area analysis of polished samples to determine mineral properties such as composition and distribution.

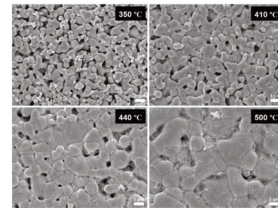


BSE image (left) and MLA compositional analysis (right) of $\text{Fe}_3\text{O}_4:\text{CaCO}_3$, illustrating microsphere porosity and demonstrating very high levels of $\text{Ca}_2\text{Fe}_2\text{O}_5$ (blue) compositional homogeneity.

J. Molinar Díaz, S. A. Samad, E. Steer, N. Neate, H. Constantin, M. T. Islam, P. D. Brown and I. Ahmed, *Materials Advances*, 2020, 1, 3539-3544.

In-situ stages

The nmRC has different in-situ stages facilitating dynamic studies and measurements of the properties of materials inside the SEM. These include the Gatan Murano-525 Heating Stage (heating from ambient up to 950 °C) enabling both imaging and the EBSD characterisation of phase transformations and grain growth to be observed in real time.



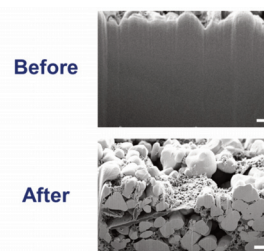
In-situ heating (350 – 500 °C) of copper nanoparticles demonstrating sintering properties.

Data courtesy of Mr Martin Roe and Dr David Pervan.

The *Deben MicroTest 200VT* stage allows both tensile and compression experiments with a load up to 200N and a variable temperature controlled platform from -20 °C to 160 °C. There is also the option to use the *MicroTest* module on the bench.

Focussed Ion Beam Scanning Electron Microscopy

FIB-SEM is a dual beam technique which incorporates the use of a secondary beam formed of gallium ions for materials processing and sample preparation including deposition, ablation and sectioning.

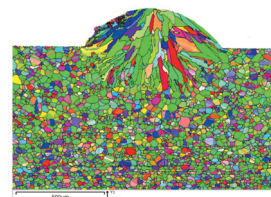


These SEM images are of Li-ion battery cathode materials before and after FIB milling using a Gallium ion beam, revealing the inner structure of the material.

Data courtesy of Mr Andrzej Sankowski.

Electron Backscatter Diffraction

EBSD is used to capture the diffraction patterns generated by the scattering of the SEM electron beam at the surface of a sample. From these patterns, the crystalline phase and the orientation of a material's structure can be determined.

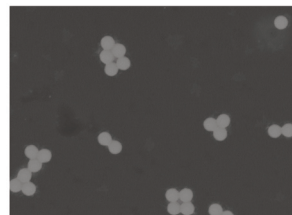


EBSD of a polished cross-section through stainless steel, showing a laser powder bed fusion (LPBF) scan track, produced by oscillated scanning laser melting. The map has inverse pole figure (IPF) colouring, to represent the orientation of the crystal structure at each point.

Data courtesy of Mr Diego Della Crociata

STEM in a SEM

A STEM detector is inserted underneath a sample inside of an SEM, in order to detect electrons, which have been transmitted through a material. Unlike TEM, the beam is focused and scanned across the sample, meaning the resulting image presents a slightly different perspective on the sample. Due to the requirement for electrons to pass through the sample, STEM requires for samples to be thin (ideally less than 200 nm).



Polystyrene nanoparticles imaged in STEM mode using the Zeiss Crossbeam 550 SEM.

Data courtesy of Ms Zoe Tanner and Dr Chris Parmenter

Find out how in-situ SEM techniques could help with your applications, designs or solutions:

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Transmission electron microscopy (TEM)

Transmission electron microscopy is an electron microscopy technique capable of imaging with a resolution down to an Ångstrom scale (~ 0.19 nm). It uses the spatial contrast generated by variations in electron transmission as they pass through specially prepared ultra-thin specimens to generate an image.

TEM can also provide advanced structural, crystallographic and chemical characterisation of samples on the nanoscale.

Capabilities

- Ultra-high magnification and resolution imaging.
- Micro- and nano-structural characterisation.
- Simultaneous elemental and compositional analysis.
- Thickness, pressure and process measurements.
- Nanotomography (3D profiling).

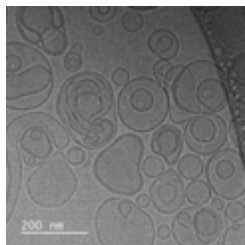
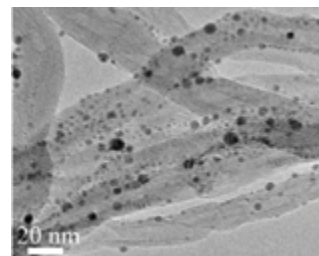
Typical applications

- Nanostructural analysis and component identification.
- Interrogation of coating, multi-phase alloy, fibre (and other) ultrastructures.
- Tissue and cellular imaging of with full-structure visualisation.
- Cryogenic visualisation of solution or suspension based nano-structures, such as liposomes.
- Small structure electron crystallography.

Imaging molecular assemblies

TEM offers nanoscale, high magnification imaging and chemical analysis. The smallest structural systems down to the atomic scale can be imaged and characterised in real time. Here silver nanoparticles can be visualised as having been successfully encapsulated in multi-walled carbon nanotube. Such capability ensures complex physicochemical processes to produce novel materials with bespoke characteristics can be validated for example size control, electron transport, heat transfer and others.

JA Watts, MW Fay, GA Rance, PD Brown, AN Khlobystovbc. *Carbon* 139 (2018), 538-544.



Electron transfer across biological membranes

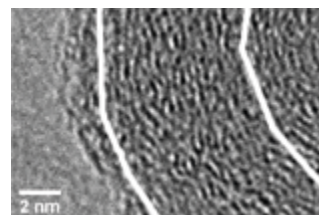
TEM was used to investigate the use of carbon nanotube porins (CNTPs) as wireless bipolar electrodes and artificial voltage-dependent anion-selective channels (switchable porins within the membrane) within biological systems. CNTPs were self-inserted into giant unilamellar vesicles and the fine-details of subsequent CNTP presence within the membrane was visualised with use of a cryogenic-TEM method.

JM Hicks, YC Yao, S Barber, N Neate, JA Watts, A Noy, FJ Rawson. *Small* 17 (2021), 2102517.

Quantifying soot nanostructures

TEM offers unique system diagnostics by combining molecular level structural and chemical information, such as particle morphology, size distribution, elemental presence/absence and others. One such example is soot-nanostructures. TEM analysis revealed the atomic structuring of the nano-sized soot-in-oil particulates and agglomerates from a gasoline direct injection deposit. This morphological information was used to evaluate image processing parameters, relevant for lattice fringe analysis, a common method to quantify soot nanostructures. White lines indicate the region of interest in this investigation.

SA Pfau, A La Rocca, MW Fay. *Combustion and Flame* 211 (2020), 430-444.



Our facilities

JEOL 2100F FEG-TEM

- Field emission electron gun (FEG) instrument, for use at 100kV and 200kV.
- A point resolution of 0.19nm.
- Bright field STEM detector.
- High angle annular dark field (HAADF) STEM detector.
- Gatan K3 IS 23.6 megapixel, electron counting direct detection (DDE) camera. Capable of 150 frames per second at full view, or >3500fps at 256x256 pixels.
- Gatan Tridiem Filter Spectrometer and 2K x 2K CCD camera, configured for use at 100kV and 200kV. Enables elemental mapping via electron energy loss spectroscopy (EELS) and energy filtered TEM (EFTEM).
- Oxford Instruments 80mm X-Max system for energy dispersive X-ray spectroscopy (EDS) analysis.
- Room temperature tomography: Gatan 916 room temperature tomography holder with up to 80 degrees tilt.
- Cryo-tomography and cryo transfer: Gatan 914 and Gatan Elsa Cryo-tomography holders including cold controller/ cryo-workstation.
- Electrical holder. Gatan 936 DT analytical LN2 holder with temperature controller with EBIC stage option (four electrical connections) plus Smart EBIC.
- Gatan 4004 heating and gas exchange holder. Allows samples to be heated up to 800 °C, or air sensitive sample analysis.

JEOL 2100+ TEM

- LaB6 TEM for high throughput, high versatility analysis at 80kV or 160kV.
- Gatan OneView camera. High-resolution, 16-megapixel CMOS camera. Capable of 25 full frames per second or 300fps at 512x512 pixels.
- Bright field STEM detector.
- Oxford Instruments X-MaxN 80 TLE EDS detector.
- Gatan Enfinium EELS detector.
- HAADF detector.
- Range of specialised sample rods including heating and cryogenic stages.
- MEMS Heating holder. DENSolutions Wildfire S3 capable of analyses up to 1300°C with millisecond heat and quench speed, and nanoscale sample drift with step changes of hundreds of degrees. Enables EELS and EDS mapping at elevated temperatures.

FEI Tecnai G2 12 Biotwin

- 120 kV LaB6 TEM for high contrast imaging.
- Gatan SIS Megaview IV digital camera.
- Tomography compustage for tomographic characterisation.
- Cryo-stage for low temperature observation of temperature dependent or hydrated samples.
- Ideal for low-contrast, beam-sensitive biological specimens, or other soft materials such as polymers.
- Gatan Orius (4k x 2.6k) Camera with digital streaming video for high-resolution and TV rate imaging.

Find out how TEM could help with your applications, designs or solutions:

nmrcenquiries@nottingham.ac.uk +44 (0)115 951 5046

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Raman spectroscopy

Raman spectroscopy is a powerful analytical tool for materials characterisation, providing key information on the structure, chemical composition and local environment of molecules using the diagnostic fingerprint the vibrational spectrum uniquely delivers.

Using the inelastic scattering of light, it enables analysis of molecular materials in their native state, in the absence of labels or complex preparation procedures, in a non-invasive and non-destructive fashion.

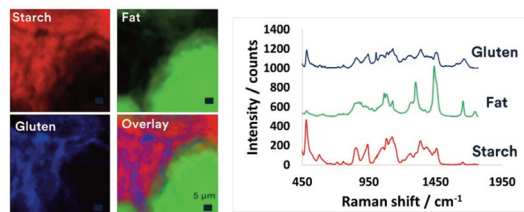
Combining confocal microscopy with automated mapping allows for the creation of false colour images in 1D, 2D, or 3D. These images use brightness, contrast, and colour to illustrate the science of the sample.

Capabilities

- Ambient chemical composition and structural analysis.
- Real-time material transitions.
- Controllable confocality for depth profiling and multi-dimensional mapping.
- Several discreet laser sources including UV, visible and NIR.
- Environmental control for real-time materials analysis in *operando*.

Typical applications

- Pharmaceutical agents and cosmetic products.
- Geology and mineralogy.
- Carbon materials.
- Semiconductors.
- Life sciences.



Raman of ramen: imaging the distribution of ingredients in foodstuffs

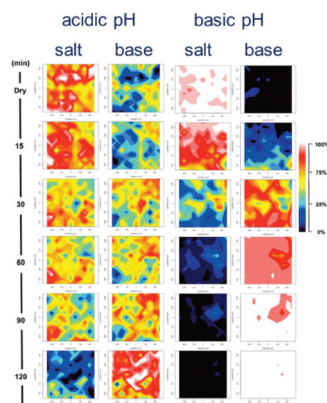
Foodstuffs consist of multiple ingredients, the shape and size of which affects fundamental properties like texture and taste. To probe the nature and distribution of ingredients within ramen noodles – a prototype foodstuff – a square area from the surface of a store-bought noodle was chemically imaged using Raman mapping. The identities of the ingredients were determined using chemometric analysis and indexing relative to previously reported spectra, with the resultant false colour images indicating that the surface of ramen noodles comprise a complex mixture of fat droplets within a matrix of flour (gluten and starch).

Data courtesy of Dr Graham Rance

Watching drugs dissolve: probing the effect of excipients on salt disproportionation in pharmaceutical tablets during dissolution

An estimated 50–70% of all small molecule drugs are administered as salts. However, their applicability in tablets significantly depends on their stability both before and during drug release, as there is a tendency for the salt to convert back to its free base form via a reaction known as salt disproportionation. Using Raman spectroscopy it is possible to discriminate the salt and free base forms and in this study we explored the effect of excipients of the pH-dependent disproportionation of the anti-diabetic medication Pioglitazone during dissolution. By applying a bespoke flow cell, we were able to simultaneously monitor the composition of the tablet by Raman mapping and the extent of drug release by UV-vis spectroscopy as the tablet dissolved.

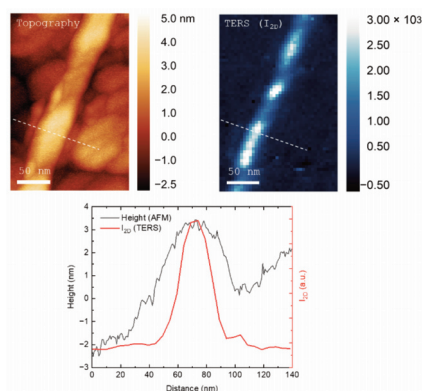
A. Abouselo et al., *Molecular Pharmaceutics*, vol. 18, no. 9, pp. 3247-3259,



Raman below the diffraction limit: imaging individual carbon nanotubes using TERS

The chemical analysis of molecular materials using Raman spectroscopy is fundamentally restricted by the optical diffraction limit (~ 650 nm spatial resolution with $\lambda=532$ nm and $N.A=1$). Addressing individual objects at the nanoscale therefore requires an alternative analytical approaches. Of these, Tip-Enhanced Raman Spectroscopy (TERS) – which combines Raman spectroscopy and scanning probe microscopy – has been receiving increased interest, and commercial TERS is now available. Here, we demonstrate the nanoscale imaging capabilities of TERS facilities at the nmRC, providing simultaneous topographic and spectroscopic molecular fingerprint of a single carbon nanotube measuring less than 5 nm in diameter.

Data courtesy of Drs Graham Rance and James Kerfoot



Our facilities

- Controllable confocality for standard or high spatial resolution hyperspectral imaging.
- Ultra-low frequency (ULF) Raman module allowing measurement in the sub-50 cm^{-1} region.
- SWIFT™ imaging capability for ultra-fast mapping applications.
- EasyNav™ software module for optimal Raman imaging.
- Multivariate analysis (MVA) software module for advanced data processing.
- KnowItAll™ database for spectral searching, analysis and data mining.
- DuoScan™ imaging system for sub-micron to macro-scale mapping.
- Multiwell software module and holder for high throughput screening using traditional well plates and related regular array sample geometries.
- ParticleFinder software module for automated location, characterisation and Raman analysis of particles.

- Linkam THMS600 variable temperature stage (-196 to 600 °C).
- Custom-designed flow cell for correlative 'in-situ' dissolution and elution studies.
- Bespoke cell for Raman spectroelectrochemistry studies.

HORIBA LabRAM HR Raman Microscope

- Upright microscope geometry.
- Excitation wavelengths available: 325, 532, 660 and 785 nm.
- Gratings available: 300, 600, 1200 and 1800 lines/mm (depending on configuration).
- Objectives available: 10x, 40x, 50x and 100x (depending on configuration).

HORIBA XploRA INV Raman Microscope

- Inverted microscope geometry, suitable for 'in-situ' analysis and biological samples.
- Excitation wavelength available: 532, 638 and 785 nm.
- Gratings available: 600, 1200, 1800 and 2400 lines/mm (depending on configuration).
- Objectives available: 10x, 20x, 50x, 60x and 100x (depending on configuration).

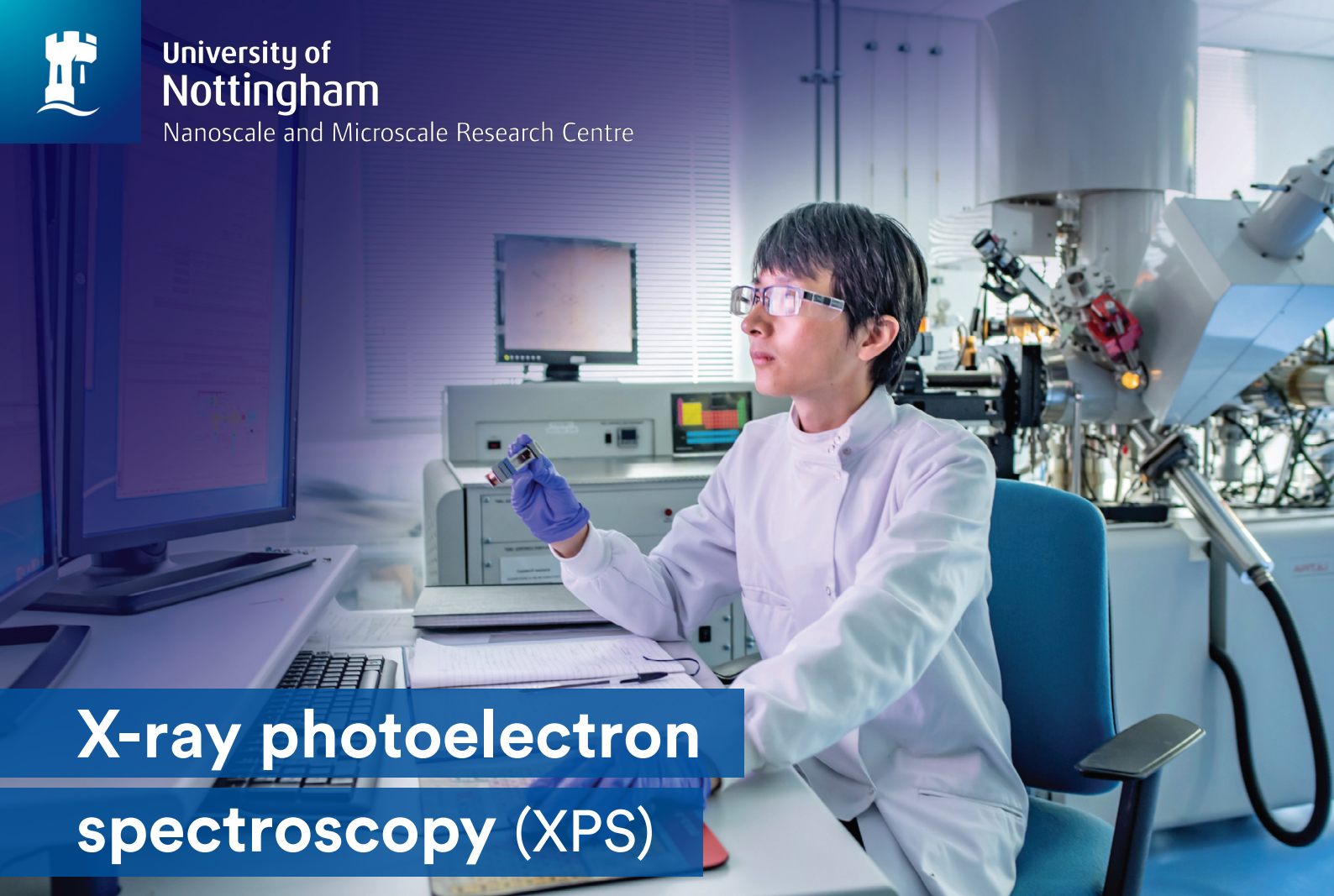
HORIBA LabRAM HR Evo Nano platform (DCI-TERS)

- Scanning probe microscope coupled to a Raman microscope enabling tip-enhanced Raman spectroscopy and co-localised AFM-Raman.
- Conventional upright microscope geometry.
- Excitation wavelengths available: 532, 633 and 785 nm.
- Gratings available: 150, 600, 1800 and 2400 lines/mm.
- Objectives available: 5x, 10x, 50x, and 100x.
- Motorised half-wave and analyser plates for polarised Raman measurements.
- AFM imaging modalities including topography (contact, semi-contact, non-contact), KPFM, SCM, EFM, PFM, cAFM, LFM, FMM, MFM and STM.
- Dual optical access from the side and below for reflectance and transmission TERS measurements.
- Holders for variable temperature analysis in air (-50 to $+300$ °C) and liquids (ambient to $+60$ °C).
- Electrochemical cell for ecTERS.

Find out how Raman spectroscopy could help with your applications, designs or solutions:

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X-ray photoelectron spectroscopy (XPS)

X-ray photoelectron spectroscopy provides a quantitative measurement of surface (5-10 nm) elemental composition and chemical state using X-ray stimulated photoelectron emission.

Characteristic electron binding energy ‘fingerprints’ for a sample can be produced, allowing determination of both atomic composition and subtle chemical state variations. Elemental imaging can be formed from XPS data, and depth profiling is a powerful adjunct to quantify progressive near-surface chemistry.

Capabilities

- Quantification of surface elemental composition
- Quantification of surface chemical state, such as $\text{Fe}^{2+}/\text{Fe}^{3+}$ or carboxyl/alcohol
- Depth profiling using an Ar gas cluster ion source
- Elemental mapping (parallel XPS imaging)

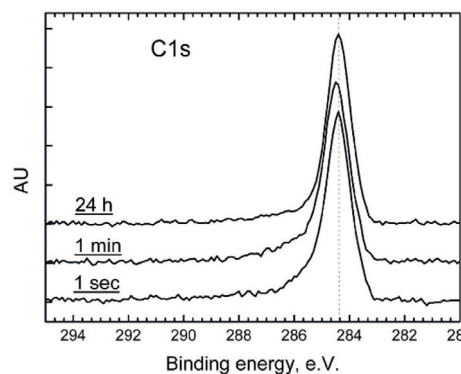
Typical applications

- Contaminant identification and quantification (stains, corrosion, residues, phase separations)
- Oxidation state analysis
- Quantification of surface chemical modifications
- Thin film thickness (up to 15 nm) and composition quantification
- Catalyst surface characterisation

Quantification and qualification of self-assembled monolayer thickness

XPS can be used to qualify and quantify surface modifications. For example high resolution XPS carbon (C1s) region scans can be used to assess alkane thiol binding to gold (Au) surfaces when prepared at different adsorption times. The quality is assessed from the C1s region, which lacks any noticeable peaks related to oxygen-containing groups or any other possible contaminations. Au Layer thickness can also be quantified from the intensity of the C-C peak compared to the Au 4f core level (not shown).

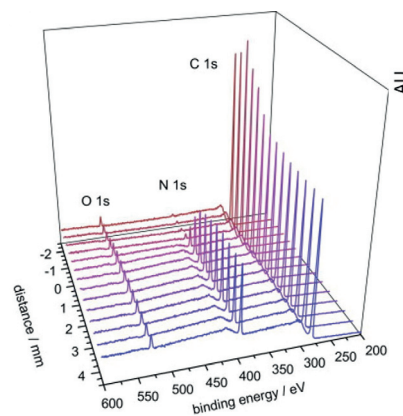
Vladimir V Korolkov, Stephanie Allen, Clive J Roberts, and Saul JB Tendler. *Journal of Physical Chemistry* 115 (2011), 14899–14906.



Quantification of chemical gradients

XPS can be used to provide characterisation of subtle differences in spatial chemical character. Here progressive (0.5mm increments) XPS widescans along a graded chemical interface are used to quantify the elemental concentration. This system is a thin hydrocarbon polymer deposited with a gradually changing thickness on a nitrogen containing polymer with uniform thickness. The gradient can be fully characterised from this information which allows the performance of this functional surface to be understood.

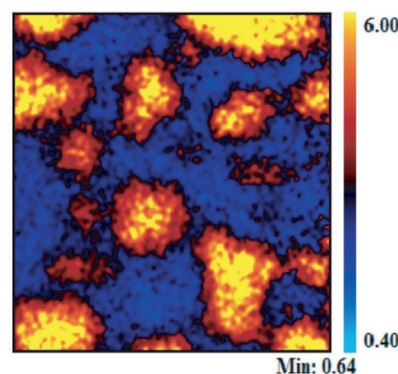
Mischa Zelzer, Ruby Majani, James W Bradley, Felicity RAJ Rose, Martyn C Davies, Morgan R Alexander. *Biomaterials* 29 (2008), 172–184.



XPS Imaging of the elemental corrosion in an optical filter

Spatially sequential XPS spectra can be processed into images to visualise elemental and chemical state distributions. This figure shows the corrosion of germanium to germanium oxide on an optical filter, as seen by XPS. An intensity map for germanium oxide (Ge 3d and O 2s) over a 400µm x 400µm area indicates significant variability incurred by moisture induced corrosion. The scale bar represents atomic %.

Emily F Smith, David Briggs, Neal Fairley. *Surface and Interface Analysis* 38 (2006), 69–75.



Our facilities

Kratos Liquid Phase Photoelectron Spectroscopy Machine (LiPPS)

- Multiple monochromated X-ray sources including Al K α emission at 1486.6 eV, Mg, or a high energy Ag source at 2984 eV.
- Argon gas cluster source for high resolution depth profiling of organic materials (biological samples and polymers).
- High throughput multi position programmable stage allows multiple samples in one experiment including a tilt stage for topographic samples.
- Magnetic immersion lens system allows the area of analysis to be defined by apertures.
- Electrostatic/magnetic lens system (hybrid lens) and a hemispherical analyser (CHA) to sort photoelectrons according to kinetic energy.
- Electron detection and counting with a triple channel plate and delay line detector (DLD).
- Heating and cooling capabilities.
- Electrochemistry stage for in situ work on ionic liquids.

Thermo Fisher K-Alpha Photoelectron Spectrometer

- Al K α emission X-ray source at 1486.8 eV
- Ion gun with energy range of 200–4000 eV
- 180° double focussing hemispherical analyser with 128-channel detector
- Dual beam source charge compensation
- 4-axis sample stage, 60 × 60 mm sample area, 20 mm maximum sample thickness

Find out how XPS could help with your applications, designs or solutions:

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Secondary ion mass spectrometry (ToF-SIMS & 3D OrbiSIMS)

Secondary ion mass spectrometry is used to characterise the surface chemistry of a material. A beam of primary ions impacts the surface and liberates secondary ions from the sample which are then analysed in turn to produce mass spectra. The lateral distribution of chemical species (mapping) or their intensity with depth (depth profiling) can then be carried out.

The combination of a time-of-flight secondary ion mass spectrometer (ToF-SIMS) with hybrid OrbiTrap™ functionality provides an array of analytical options to provide 3D chemical analysis with exceptional surface sensitivity (1-3 nm), high mass and spatial resolutions.

Capabilities

- Label-free large molecule identification ($> m/z$ 1000)
- Sensitive trace element identification (ppm)
- Chemical mapping (down to a nm scale)
- Depth profiling of inorganics and organics

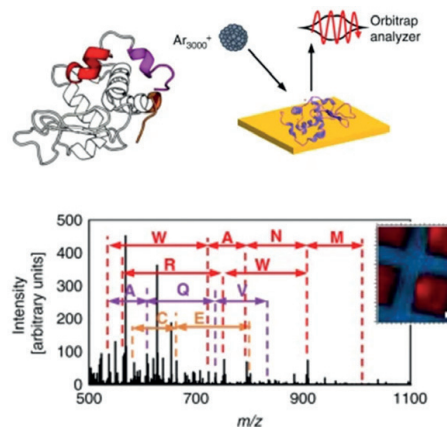
Typical applications

- Identification of unknown organic species in solids
- Contaminant identification and distribution
- Surface (such as coatings, films, deposits) composition and integrity
- 3D permeation assessment of active pharmaceutical ingredients
- Spatial resolution of chemical components.
- High throughput screening of polymers

In-situ protein identification for next generation biomaterials and tissue analysis

Using the ballistic fragmentation and high accuracy of the 3D OrbiSIMS, 16 undigested proteins were identified in-situ (in their native state). This was achieved without a chemical label, enzymatic digestion or use of a specific matrix. Using the ballistic approach the concentration of key proteins was tracked into human skin. The ability to directly measure proteins in their native state using a surface analysis technique has significant potential application in furthering the understanding of diseases and the development of new bio-materials.

AM Kotowska, GF Trindade, PM Mendes et al. Protein identification by 3D OrbiSIMS to facilitate in situ imaging and depth profiling. *Nature Communications* 11 (2020), 5832. doi.org/10.1038/s41467-020-19445-x

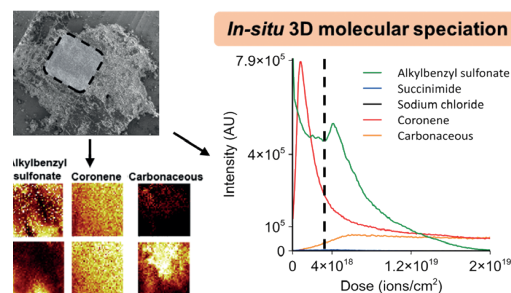


3D insight into the molecular composition and formation of polluting engine deposits

Formation of deposits in internal combustion engines causes increased emissions and lower engine efficiency. Using the 3D OrbiSIMS technique it was possible to depth profile and image these complex layered materials and, coupled with sophisticated chemical filtering using molecular formula prediction for data processing, a comprehensive molecular characterisation of petroleum deposits was performed. Argon gas cluster depth profiles tracked the fate of molecules once deposited on the engine surface and unveiled plausible formation pathways of deposits for the first time. The combined insight into the composition, origin and formation of deposits will help mitigate the use of solubilising fuel additives and help reduce worldwide vehicle emissions.

M Edney, J Lamb, M Spanu, E Smith, E Steer, E Wilmot, J Reid, J Barker, M Alexander, C Snape and D Scurr. *ACS Applied Materials and Interfaces* 12 (2020), 51026–51035.

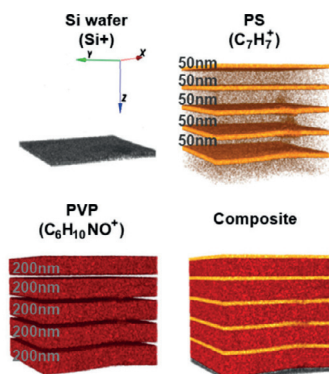
MK Edney, AM Kotowska, M Spanu, GF Trindade, E Wilmot, J Reid, J Barker, JW Aylott, AG Shard, MR Alexander, CE Snape, DJ Scurr. *Analytical Chemistry* 94 (2022), 4703–4711.



3D ToF-SIMS imaging of polymer multi-layer films

ToF-SIMS imaging with argon cluster sputter depth profiling can provide a detailed insight into the three-dimensional chemical composition of organic material structures. 3D chemical images can provide information regarding the structure of multi-layer systems which can be used to inform future manufacturing and development. Outputs include sample layer chemistry, homogeneity, thickness and interface widths. Here we can see the analysis of spin-cast multi-layers comprising alternating polystyrene (PS) and polyvinylpyrrolidone (PVP) layers. The quality of the data allows a detailed analysis of the chemical structure of these systems, revealing minor imperfections within the polymer layers.

J Bailey, R Havelund, JS Sharp, AG Shard, I Gilmore, MR Alexander, DJ Scurr. *ACS Applied Materials and Interfaces* 7 (2015), 2654–2659.



Our facilities

ION-TOF (GmbH) ToF SIMS V

- Liquid metal (Bin+n) ion gun (LMIG) for spectroscopy and imaging at a spatial resolution of ~ 200 nm.
- Argon gas cluster source for high-resolution depth profiling of organic materials (polymers and biological samples) and 3D chemical characterisation.
- Mass sensitivity down to ppm (femtomole).
- A 5-axis multi-sample stage is fully automated and provides rotation for high-resolution (nm) depth profiling (Cs+ or Ar GCIB sources).
- Reflectron ToF mass analyser gives mass resolution > 13000 at m/z = 29.
- Chemical imaging of surface areas from the µm to cm scale.
- 3D elemental mapping possible.
- Sample size accommodation from a few mm up to ~ 10 cm.

Ion-TOF (GmbH) HybridSIMS

- ToF or Orbitrap™ analysis of organic and inorganic samples including petroleum deposits, biological materials (protein identification, skin, hair, leaves and others), polymers, semiconductors, insulators, powders, foils and microarrays.
- High mass resolution spectrometry (>240,000 and 11,000 amu for the Orbitrap and the ToF, respectively).
- High spatial resolution chemical imaging (<70 nm).
- Mass sensitivity down to ppm (femtomole).
- Gas cluster ion beam sputtering for controlled depth profile analysis of organics.
- Biosafety Level 2 (BSL2) preparation and analysis environment for cell/tissue analysis including cryogenic sample preparation facility, including high pressure freezing, freeze drying, cryo-ultramicrotomy and more.
- Chemical filtering and multivariate analysis software and expertise for complex chemometric data analysis.

Find out how ToF/3D OrbiSIMS could help with your applications, designs or solutions:

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A photograph of a female scientist with curly brown hair and glasses, wearing a white lab coat, looking through the eyepiece of a confocal microscope. The microscope is a large, complex piece of equipment with various lenses and a stage. In the background, there are computer monitors displaying colorful, abstract images, likely representing microscopic data. The scene is set in a laboratory with blue and green lighting.

Confocal microscopy

Confocal laser scanning microscopy (CLSM) is a state-of-the-art imaging technique that utilises laser light to scan and create high-resolution, three-dimensional images of biological and non-biological specimens. This method allows for precise visualisation of fine structures and selectively imaging specific depths within a sample, resulting in improved analysis of cellular and molecular processes.

Capabilities

- Structural properties of cells by fluorescence.
- Live cell imaging.
- 3D Z-stacks.
- Biofluidic microscopy.
- Correlative light and electron microscopy.

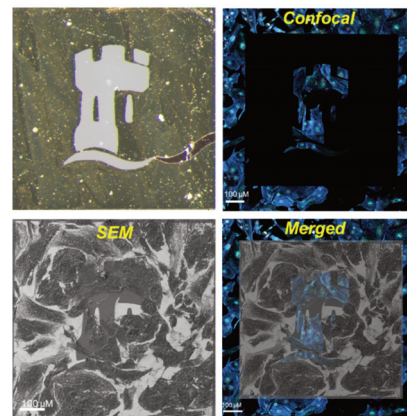
Typical applications

- Animal cells and tissues.
- Microbiology and biofilm research.
- Plant and crop science.
- Advanced materials, bioengineering, pharma and foods.

Correlative confocal and electron microscopy of cells grown on a substrate

To demonstrate the correlative nature of confocal and electron microscopy, photolithography was used to pattern gold onto a coverslip with the UoN logo as the design. STEM cells were then grown on top and stained with fluorophores to image actin (cellular skeleton), mitochondria, and the nucleus. Imaging using confocal microscopy was able to demonstrate the cell structure and components, whilst electron microscopy revealed the topography of the sample. When the two images are merged, a holistic view of the sample could be obtained.

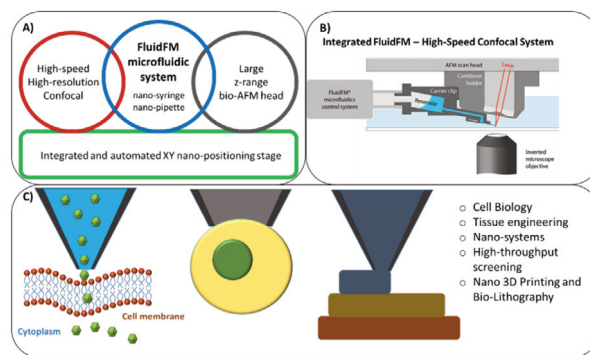
Data courtesy of Drs Jacqueline Swift, Aishah Nasir, Kate Nguyen, and Richard Cousins



Biofluidic Microscope (BFM) setup at the nmRC

The BFM is a new integrated imaging platform that combines ultra-fast confocal imaging with the atomic force and nano-fluidic functionality to enable characterisation of localised biochemical and physiological processes. A) Conceptual diagram of key components of the BFM that brings together AFM/FluidFM analysis, sample positioning, ultra-fast optical detection, as well as novel approaches for image processing. B) Schematic diagram of the BFM operating in a FluidFM mode. C) Some key novel applications of the system.

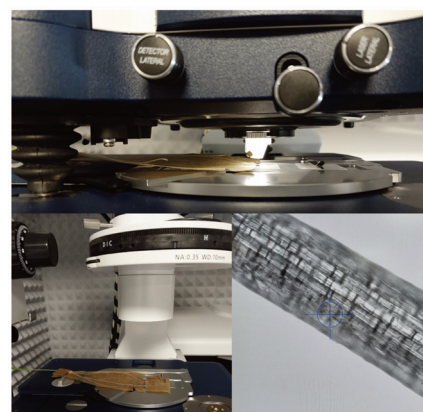
Images and schematics courtesy of Dr Gleb Yakubov



Imaging of large and live barley shoots

Barley shoots (kept live) are placed under a microscope as a whole object without cutting or dissecting. The use of the customised platform enables to perform these tests under water to keep the roots moist. Visualisation was performed using both bottom and top view optics. The task was to probe mechanical properties of cells as a function of their distance from the root tip. This was performed using Force Volume Scanning using CellHesion Head to enable tracking large variations in topography of cells arranged around a cylindrical morphology of the root.

Images and data courtesy of Dr Gleb Yakubov



Our facilities

Two Zeiss 900 CLSMs coupled with Airyscan 2.

- Inverted and upright geometries available
- Super-resolution of 140 nm
- 32 circularly arranged detection elements
- Zeiss PALM MicroTweezers – optical tweezers system that allows precise, contact-free cell manipulation
- Linkam CMS196 Cryo-Correlative microscopy stage allowing cryo-confocal imaging
- 3D Z-stack viewing
- Incubator stage for live imaging
- Zen Blue software for flexible workflows

Biofluidic Microscope

- Zeiss LSM 980 microscope Airyscan 2 with Multiplex Mode and maximum resolution of 120 nm
- Bruker NanoWizard V Scanning Head
- JPK CellHesion Head with 100 µm Z-range for large and strongly adhesive samples
- CytoSurge Fluid Force Microscope (FluidFM) which uses microsyringe, micropipette, and micro-trap FluidFM cantilevers
- Double-View optics which enables reflected and transmitted light to be applied simultaneously.
- PetriDish Heater – versatile holder for a wide range of 35 mm Petri dishes, including glass-bottom dishes. Heats up to 60 °C

Find out how Confocal microscopy could help with your applications, designs or solutions:
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Cryogenic materials characterisation

Cryogenic materials characterisation refers to the use of low temperatures to enable the measurement and observation of the physical and chemical properties of materials. These techniques are particularly useful for studying hydrated material, biological specimens or volatile samples to retain them in their native state. These species can be difficult to study using conventional characterisation techniques as they are sensitive to damage by vacuum conditions or exposure to an electron or ion beam. Cryogenic techniques can also be used for time series studies or to stabilise samples during imaging.

Capabilities

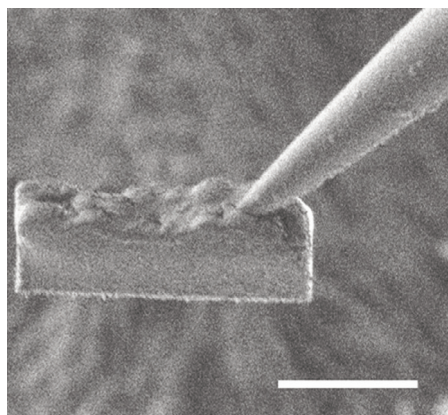
- Imaging and spectroscopy of frozen and soft matter materials
- Tomographic measurements of frozen and soft matter materials
- Cryo-focussed ion beam scanning electron microscopy (Cryo-FIB-SEM)
- Cryo lift-out of thin sections
- Detection, imaging or distribution mapping of volatile compounds
- Analysis of biomolecules and biological samples in their native state – maintaining hydrated state of samples such as hydrogels, tissues, biofilms

Typical applications

- Micro and nanoscale structural, compositional, and cross-sectional determination of biological and high-water content materials
- Sensitive trace element identification (ppm) and nanoscale chemical mapping
- Studies of frozen material using correlative techniques

Cryo-FIB-lift-out: practically impossible to practical reality

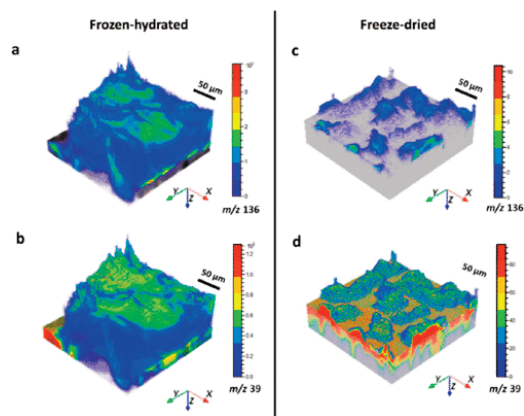
Cryo lift-out allows the preparation of lamella structures from a bulk sample by initially cutting through a sample using focussed ion beam (FIB) SEM and then using a cooled micromanipulator to secure the thin specimen. These lamellae can then be transferred for Cryo transmission electron microscopy imaging for further analysis. Shown is an overview of the Cryo lift-out procedure including; approach and positioning of the micromanipulator, lamella contact, attachment of the micromanipulator, milling of connection between lamella and the sample, and successful securement of the lamella to the tip.



C. D. Parmenter and Z. A. Nizamudeen. *Journal of Microscopy* 281 (2020), 157-174.

Cryo-OrbiSIMS for 3D molecular imaging of a bacterial biofilm in its native state

Cryogenic conditions allow analysis of bacterial biofilms in a native state and mapping of the chemistry for a highly hydrated sample in 3D. Samples composed mainly of water lose their 3D structure when drying. Shown below is a comparison of ToF-SIMS images of (a, b) the frozen hydrated biofilm and (c, d) freeze-dried biofilm. The samples were prepared using a Leica EM ICE high-pressure freezer and transported to the instrument using a Leica EM Vacuum Cryo Transfer system.



J. Zhang, J. Brown, D. J. Scurr, A. Bullen, K. MacLellan-Gibson, P. Williams, M. R. Alexander, K. R. Hardie, I. S. Gilmore, and P. D. Rakowska. *Analytical Chemistry* 92 (2020), 9008-9015.

Our facilities

Instrumentation

JEOL 2100F Transmission Electron Microscope (TEM)

High resolution TEM with a direct detection electron camera, and EDS capabilities.

JEOL 2100+ TEM

TEM with a high performance camera and EDS capabilities.

Thermo Fisher (FEI)

Tecnai G2 12 Biotwin TEM

TEM suited towards biological samples.

Thermo Fisher (FEI)

Quanta2003D Dual Beam FIB-SEM

Focussed ion beam SEM used for milling samples.

ZEISS Crossbeam 550 SEM

Focussed ion beam SEM with EDS, EBSD, and STEM capabilities

Sample preparation equipment

Leica EM GP2 and Gatan CP3 Automatic Plunge Freezers

Plunge freezing of liquid or thin samples into liquid ethane with automatic blotting.

Gatan Cryo 626 single tilt holder

Conventional low temperature TEM imaging.

Gatan Cryo 914 high tilt holder

Low temperature transfer and tomographic TEM studies.

Gatan ELSA Cryo-transfer holder

Frost-free transfer into a TEM.

Leica EM GP2 or Gatan CP3 Automatic Plunge Freezer

Plunge freezing of liquid or thin samples into liquid ethane with automatic blotting.

Leica EM ICE High pressure freezer

Immobilisation of aqueous samples at simultaneously low temperature and high pressure.

Quorum 3010 preparation system

Cryogenic preparation chamber allowing fracture, sublimation and coating (platinum) under vacuum.

Quorum 3010 Cryo rotate stage

Rotating stage maintaining sample at cryo-temperatures and permitting full range of movement in the FIB-SEM.

Linkam CMS196 v1 and v3 Cryo-Correlative Microscopy Stage

Cryogenic stage for light and fluorescence microscopy or confocal microscopy. Includes a transfer system for up to three frozen TEM grids, or three high pressure frozen 3mm planchettes to enable correlative light and electron microscopy (Cryo-CLEM). Can be used for larger samples, 6mm planchettes, 1cm coverslips, on request. Can be used to freeze samples in situ, if ultrafine structural preservation is not required.

Leica MM80 Metal Mirror Freezer

Suitable for vitreous freezing of (approx. 15µm) thin preparation of gels, creams, pastes, tissue.

3D OrbiSIMS

Label free large molecule identification.

As above

Leica EM VCT and VCM

Transfer of samples under cryogenic conditions.

Find out how cryogenic techniques could help with your applications, designs or solutions:

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A woman with blonde hair, wearing a dark red patterned sweater, is seated at a desk in a laboratory. She is looking at a computer monitor displaying a colorful AFM image. To her left is a Bruker FastScan AFM system with a protective glass enclosure. The background shows laboratory shelves with various items.

Atomic force microscopy (AFM)

Atomic force microscopy is a high-resolution scanning probe technique that allows visualisation of material surfaces and their topographical features with nanometre resolution.

The interaction between a probe tip and the surface is measured via its deflection in response to surface properties. Variations of the technique allow you to calculate surface physicochemical and mechanical properties, measure interaction forces between material surfaces and map the thermal or electrical properties of a sample.

Capabilities

- Micro- and nano-structural visualisation.
- Texture analysis (roughness, topography, morphology).
- Surface forces quantification (adhesion, surface free energy and others).
- Surface mechanical properties assessment (hardness, Young's Modulus, friction and others).
- Mapping of different surface components and phases.
- Biomolecular interactions.

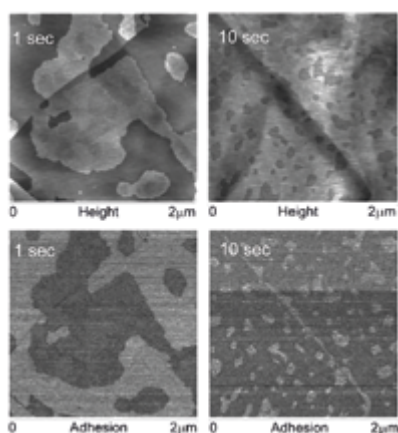
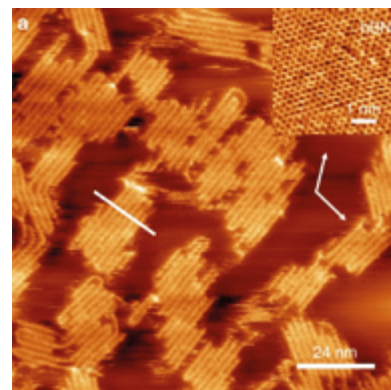
Typical applications

- Identification and localisation of phases in formulations.
- Material coating assessment.
- Visualisation of cell ultrastructure and molecular dynamics.
- Quantification of ligand-receptor interactions.
- Component integrity investigation.

High-resolution AFM images of polythiophene strands adsorbed on the surface of hBN

AFM can image material surfaces with nanometre resolution. In addition to nanostructures (such as peptides, nanoparticles and nanotubes) the technique can resolve molecular/atomic structures. This figure shows individual thiophene units and where a lattice of semicrystalline spin coated films of polythiophenes (PTs) may be resolved using AFM. Real-space images of polymers with sub-molecular resolution could provide valuable insights into the relationship between morphology and functionality of novel polymer based electronic devices.

Vladimir V Korolkov, A Summerfield, A Murphy et al. Ultra-high resolution imaging of thin films and single strands of polythiophene using atomic force microscopy. *Nature Communications* 10 (2019), 1537.



Topography and adhesion characterisation of self-assembled monolayer formation

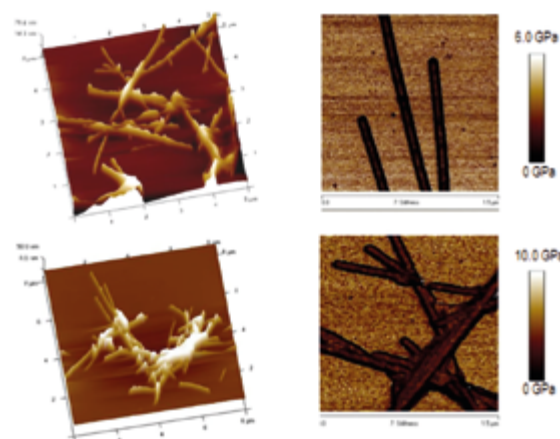
AFM allows the mapping of the adhesive character of a surface or its components. Adhesion forces can be used to calculate the interactivity of a surface and the data can be mapped to complement a topographical assessment. This figure shows height (upper row) and adhesion (lower row) AFM images (2 μm × 2 μm) detailing a real-time surface evolution due to the self-assembly of trimesic acid on a highly ordered pyrolytic graphite (HOPG) surface. The dark regions on the height and adhesion maps correspond to the areas with lower height and adhesion respectively.

Vladimir V Korolkov, Stephanie Allen, Clive J Roberts, and Saul JB Tendler. *Journal of Physical Chemistry C* 116 (2012), 11519–11525.

Mechanical mapping of peptide nanotubes

AFM can provide a mechanical assessment of a surface and its features. Here topography (3D) (left) and mechanical stiffness maps (right) are shown of nanotube structures which form in samples comprising 80% (top) and 60% (bottom) dinaphthylalanine (di-Nal) peptides to diphenylalanine peptides (FF). This combination of data channels allows functional, structural, and mechanical analyses where systems with a uniform topography but different material phases can be distinguished.

Victoria L Sedman, Xinyong Chen, Stephanie Allen, Clive J Roberts and Saul JB Tendler. *Journal of Microscopy* 249 (2013), 165–172.



Our facilities

Dimension FastScan Bio™ (Bruker): Capable of scan speeds 100 times faster than traditional AFMs, this AFM offers high-resolution, live-sample observation of interacting molecules, membrane proteins, DNA-protein binding, inter-cellular signalling and other dynamic biological processes. Includes Bruker's PeakForce™ QNM™ (Quantitative Nanomechanical Measurement) capability.

Multimode® 8 Scanning Probe Microscope (Bruker): Nanoscale imaging capabilities supplemented by simultaneous high-resolution force mapping with Bruker's PeakForce™ QNM™.

EnviroScope™ AFM (Bruker): AFM analysis within an environmental chamber capable of precise temperature and humidity control.

Many more... Including Dimension® 3100 and 3000 AFMs (Bruker), ForceRobot® 300 (JPK Instruments), MFP-1D and MFP-3D™ (Asylum Research).

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Nanofabrication suite

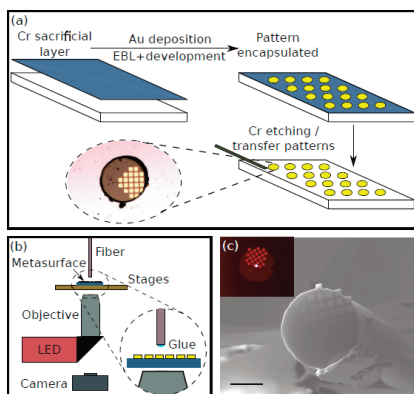
Nanofabrication is the process of creating nanometre sized features on a range of substrates. Electron beam lithography (EBL) can be used to create patterns as small as 40 nm. These patterns can then be transferred to a substrate via deposition of metals and/or dielectrics, or by selective etching of material. Our Nottingham Nanofabrication Suite can fabricate custom devices for a range of applications, and then validate them via a range of optical or thickness measurements such as ellipsometry.

Capabilities

- High resolution electron beam lithography (40 nm resolution)
- Photolithography (2 μm resolution)
- Thin film deposition of metals and dielectrics
- Wet or dry etching of substrates
- Thermal processing of materials
- Imaging ellipsometry
- Cell patterning

Typical applications

- Precise deposition of metals onto substrates such as glass
- Etching of substrates to create features such as gratings or waveguides
- Fabrication of bespoke designs for microfluidic chips
- Measurement of thickness and optical properties of 2D materials



Nanofabrication of ultrathin metallic metasurfaces for advanced imaging applications

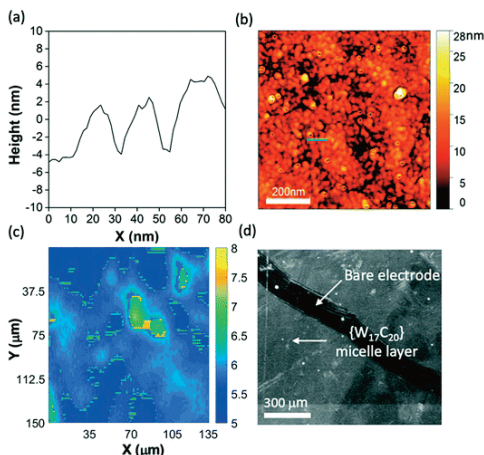
Nanofabrication can produce precise nanostructures on a surface for a range of applications. Here, a three step procedure was performed to produce ultrathin metallic metasurfaces on an optical fibre surface. Electron beam lithography was used to pattern a chrome coated silicon substrate, photolithography to pattern a layer of resist material into discs (to encapsulate the metasurfaces), and wet etching to remove the encapsulated discs. These discs were then glued onto the tip of an optical fibre for use as endoscopic devices.

Rafael Fuentes-Dominguez, Fei He, Richard B. Cousins, Christopher J. Mellor, and George S. D. Gordon. *Proceedings Volume 11953, Optical Fibers and Sensors for Medical Diagnostics, Treatment and Environmental Applications XXII* (2022), 119530F.

Imaging ellipsometry for thickness measurements of self-assembled hybrid polyoxometalate nanostructures

Using imaging ellipsometry it was possible to determine the thickness and optical properties of monolayers formed on glassy carbon electrodes by self-assembly of hybrid polyoxometalate micelles. The combination of imaging ellipsometry with complementary techniques such as atomic force microscopy (AFM) and scanning electron microscopy (SEM) allowed the 6-8 nm monolayer thickness to be determined as well as full structural characterisation of the monolayers formed that have applications in electrocatalysis and sensing.

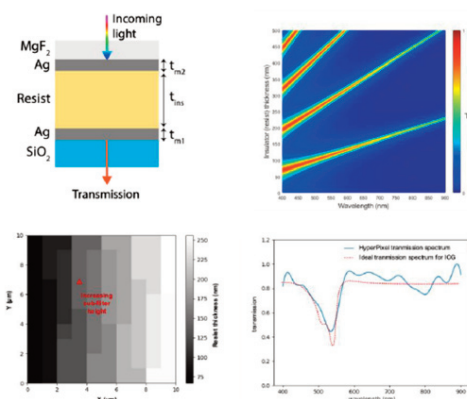
Sharad S. Amin, Jamie M. Cameron, Richard B. Cousins, James Wrigley, Letizia Liirò-Peluso, Victor Sans, Darren A. Walsh, and Graham N. Newton. *Inorganic Chemistry Frontiers* 9 (2022), 1777-1784.



Electron beam lithography for production of hyperpixel filter arrays

Electron beam lithography allows for the manufacture of unique materials that have tailored optical and electronic properties. Glass cover slips coated with a 22 nm silver layer were spin-coated with a greyscale polymer resist and then subjected to electron beam lithography to produce pixels of 0.5 – 10 μm size. The height of the pixels was also varied by controlling the electron dose. Once coated with a further silver layer this produced a filter array capable of customised spectral transmission properties.

Michaela Taylor-Williams, Richard B. Cousins, Calum Williams, Sarah E. Bohndiek, Christopher J. Mellor, and George S. D. Gordon. *Proceedings Volume 11954, Optical Biopsy XX: Toward Real-Time Spectroscopic Imaging and Diagnosis* (2022), 1195406.



Our facilities

Class 5 cleanrooms allowing for fabrication of a range of devices without the risk of contamination

Nanobeam nB5 instrument utilising an 80 kV electron beam with variable current allowing not only quick write speeds but also high resolution. Substrates from 5 to 76 mm can be used.

MJB3 and MA-6 Gen 3 mask aligners for repeated designs.

Corail 200II plasma etcher capable of reactive ion etching (RIE) and inductively coupled plasma (ICP).

Chlorinated and fluorinated gas chemistries allow for etching of Si, SiO₂, GaAs, Al, photoresist and many more materials.

Range of deposition tools allowing for deposition of metals and dielectrics

Accurion EP4 imaging ellipsometer capable of measuring optical properties and thickness of thin films with a lateral resolution of <5 μm .

Woollam M2000 variable angle spectroscopic ellipsometer (VASE) capable of making optical measurements between wavelengths of 190-1700 nm.

Alvéole Primo system attached to a **Leica DCiM inverted microscope** for cell patterning.

Find out how nanofabrication could help with your applications, designs or solutions:

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nottingham.ac.uk/nmrc or nottingham.ac.uk/nanofabrication

A photograph of a laboratory instrument, likely an isothermal titration calorimeter (ITC), with a blue-lit sample compartment. Two glass bottles with black caps and tubes are connected to the instrument. A computer monitor and keyboard are visible in the background.

Biophysical analysis

Biophysical analysis encompasses a range of techniques including Isothermal Titration Calorimetry (ITC) and Surface Plasmon Resonance (SPR). These techniques study the binding interactions of a range of biomolecules such as proteins, peptides, nucleic acids (DNA, RNA), carbohydrates, lipids as well as interactions with small molecules, polymers, ions, or drug compounds. They also obtain data on the affinity, kinetics, stoichiometry, and thermodynamics of such interactions.

Capabilities

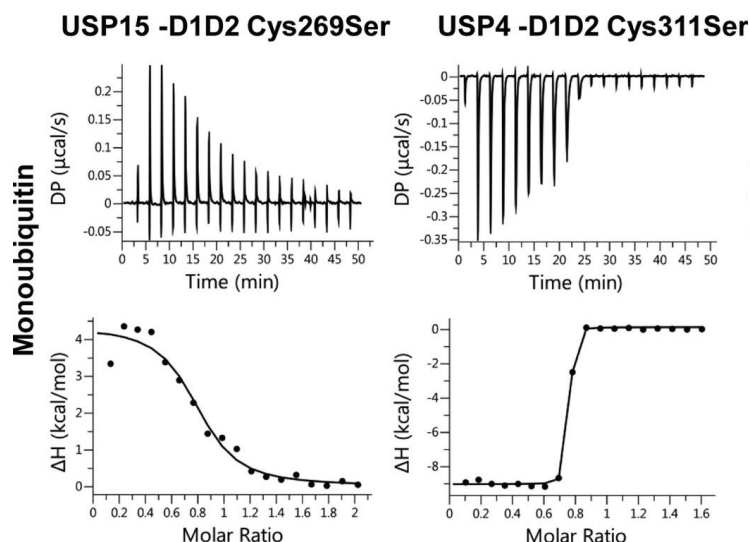
- Determination of thermodynamic parameters including binding affinity and stoichiometry
- Measurement of enzyme kinetics
- Molecular variant comparisons, e.g., mutant versus wild types
- Protein activity and stability analysis
- Epitope mapping

Typical applications

- Pharmaceutical drug discovery
- Identification of binding partners to targets
- Quality control for pharmaceuticals
- Detect and characterise molecular interactions

Enzyme-protein substrate and product interaction probed using MicroCal PEAQ ITC

ITC can enhance our understanding of the specificity of enzymes for substrates and thus was used to measure the interaction of USP15 and USP4 mutants with monoubiquitin (product) and linear diubiquitin (substrate) revealing insights on the interaction mechanism. Significant differences were observed in their thermodynamic isothermal profiles: an endothermic process was observed for the binding of USP15 mutant to both monoubiquitin and diubiquitin whereas an exothermic process was observed for USP4 mutant binding to the two proteins at 25 °C. The affinity of USP15 mutant for monoubiquitin was determined to be lower than that of USP4 mutant indicating that product inhibition plays a larger role for USP4 than USP15. Both mutant proteases had similar affinity for diubiquitin, but the interaction was associated with different enthalpy and entropy parameters.



Stephanie J. Ward, Hayley E. Gratton, Peni Indrayudha, Camille Michavila, Rishov Mukhopadhyay, Sigrun K. Maurer, Simon G. Caulton, Jonas Emsley, and Ingrid Dreveny. *J. Biol. Chem.* (2018) 293(45) 17362–17374.

SPR theory and setup

Surface Plasmon Resonance (SPR) uses optical biosensing for real-time monitoring of macromolecular interactions. In a standard SPR set up, one interacting partner (the ligand) is immobilized onto a gold sensor chip surface and an unbound interactant (the analyte) is then flowed over the surface. A change in the refractive index at the surface of the sensor (e.g., due to analyte binding or dissociation occurring near the surface) may be monitored as a shift in the resonance angle and is recorded as a sensorgram.

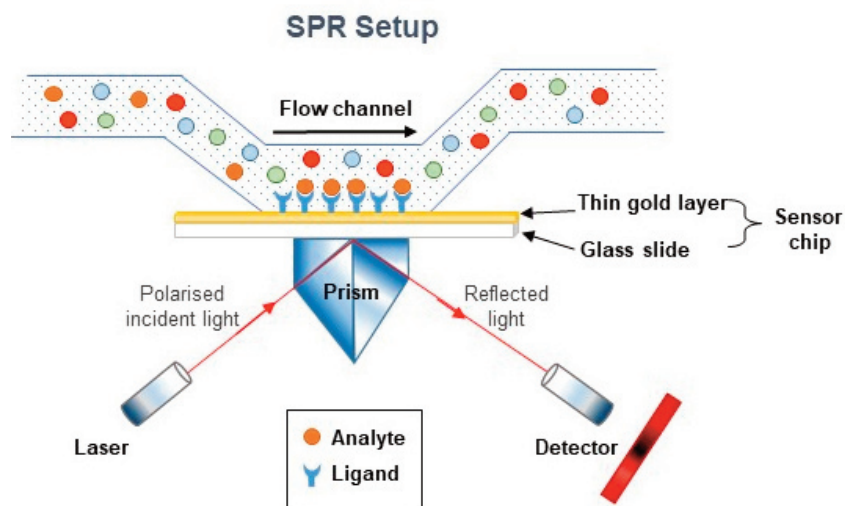


Figure courtesy of Marion J. Limo, nmRC, University of Nottingham

Our facilities

MicroCal PEAQ-ITC (Malvern)

A highly sensitive instrument designed for ease-of-use, requires as little as 10 μg of sample, and has a temperature range of 2 to 80 °C. It features user-friendly guided workflows with videos on running of experiments and performs automated cleaning of the sample cell and the titration syringe.

MicroCal PEAQ-ITC analysis software

Experimental design simulation software to guide users in selecting measurement parameters and simplify analyses with batch evaluation of large data sets and automated assessment of data quality. Final figures and graphs can also be generated quickly and easily.

Biacore T200 (Cytiva)

The Biacore T200 is a highly sensitive SPR instrument with high-throughput capabilities (up to 384 samples per run). Analysis temperature range from 4 to 45 °C, injection volume of 2 to 350 μL, affinity range from fM to mM, and concentration range > 1 pM.

Find out how biophysical analysis could help with your applications, designs or solutions:

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Particle sizing suite

Particle sizing is the characterisation of the size distribution (size range and/or mean size) and number of particles in a sample. It can be applied to solid materials, suspensions, emulsions and aerosols. Techniques include laser diffraction (LD), dynamic light scattering (DLS), nanoparticle tracking analysis (NTA), Taylor dispersion analysis (TDA), differential sedimentation (disc centrifuge (DC)) and simpler sieving and separation methods. Method selection will depend on the size range of the particles, the nature of the sample, the capabilities and limitations of the analytical method, the information and the sample throughput desired.

Capabilities

- Particle size determination
- Solution / suspension concentration
- Aggregate detection
- Assessment of colloidal stability
- Zeta potential analysis

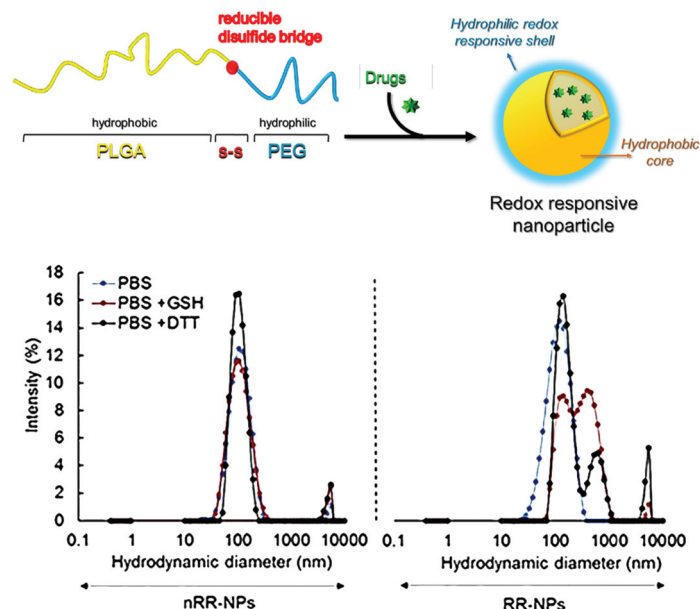
Typical applications

- Product performance; Quality control in different industries (i.e. pharmaceutical, chemical, food and energy)
- Process performance; Determination of efficiency of manufacturing process (i.e. where milling or grinding is used)
- Research and development (i.e. nanoparticle characterisation studies, study of surface modifications)

Size and zeta potential characterisation of nanoparticles designed for drug delivery

An important step in the design of drug carrier nanoparticles is characterisation of particle size and surface change in appropriate environments. Redox-responsive nanoparticles (RR-NPs) were synthesised for drug delivery into lung cancer tumour cells. The RR-NPs were designed to change surface properties when entering tumour microenvironments, which would in turn enhance their cell internalisation and delivery of drug cargo. Characterisation using a Zetasizer Nano ZS instrument of both RR-NPs and non-RR-NPs showed similar properties including hydrodynamic diameter (120 nm), low polydispersity, and high negative zeta potential values. However, size distribution curves showed lower colloidal stability of RR-NPs under in vitro reducing conditions.

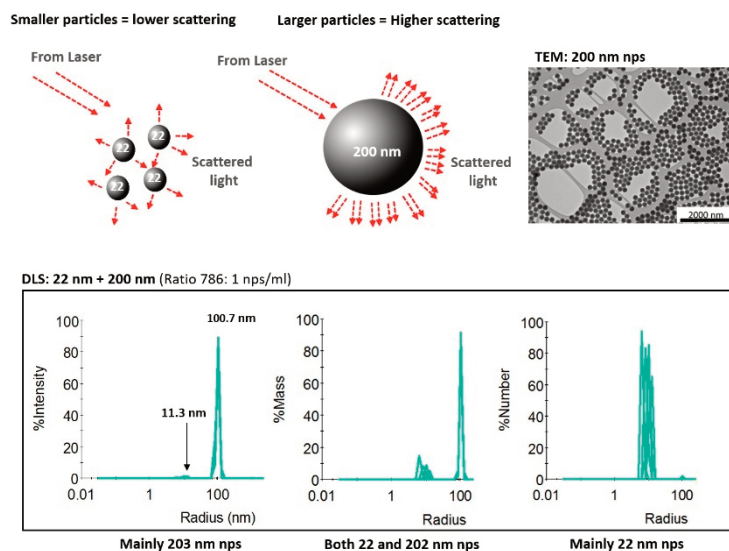
Claudia Conte, Francesca Mastrotto, Vincenzo Taresco, Aleksandra Tchoryk, Fabiana Quaglia, Snjezana Stolnik, and Cameron Alexander. *Journal of Controlled Release*, 277 (2018), 35-45.



Size analysis of a mixture of polystyrene nanoparticles

DLS measurements provide an intensity distribution of particle sizes that can also be converted to volume and number distributions. Comparing these distributions helps understand mixed populations, where larger particles scatter more light and cause variations in the distributions. A DynaPro Plate Reader II instrument was used to characterize a mixture of 22 and 200 nm polystyrene nanoparticles and found from the number distribution the 22 nm to be the major population in the mixture.

Data courtesy of Dr Marion Limo, nmRC, Nottingham.



Our facilities

DynaPro Plate Reader II dynamic light scattering instrument capable of measuring between 1 nm to 2 µm hydrodynamic diameter and providing information of polydispersity.

Zetasizer Nano ZS for electrophoretic mobility of proteins, zeta potential of nanoparticles colloids.

Viscotek 802 dynamic light scattering which can be operated between 4 and 60 °C.

LA-960 Laser Particle Size Analyser that measures sizes between 10 nm and 5000 µm from both wet and dry samples such as powders, gels, and creams.

Zetaview NTA capable of measuring hydrodynamic particle size, zeta potential, concentration, and fluorescence.

Electron Microscopy suite including scanning and transmission electron microscopy for microscale and nanoscale particle sizing down to 1-2 nm sized features.

Find out how particle analysis could help with your applications, designs or solutions:

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Liquid Chromatography- Mass Spectrometry (LC-MS)

LC-MS is a versatile and highly sensitive analytical technique for the measurement of small molecular weight compounds in a diverse range of sample types. It uses a series of mass detection systems to provide both quantitative and qualitative analyses. Situated in the Centre for Analytical Bioscience within the School of Pharmacy, our MS facilities are ideal for small molecule analysis ($<2000m/z$) and are complemented by a suite of UHPLC or specialised surface analysis interfaces (DESI, LESA) and software for data interpretation.

Capabilities

- Ultra-high-performance liquid chromatography (UHPLC) coupled to either high resolution-MS or QTrap MS.
- Accurate mass measurement (<5 ppm) for untargeted metabolomics and high resolution targeted quantitative analysis.
- Multiple reaction monitoring (MRM) and parallel reaction monitoring (PRM) for high sensitivity or specific quantitative analysis.
- Multi-stage MS and data-dependant MS fragmentation for structural elucidation and MS/MS identification.
- Ambient analysis of chemical compounds on biological/material surfaces using LESA-MS.
- Direct infusion high throughput analysis with NanoMate (no LC separation).
- Spot analysis or imaging with AP-MALDI ($5\ \mu m$ resolution).

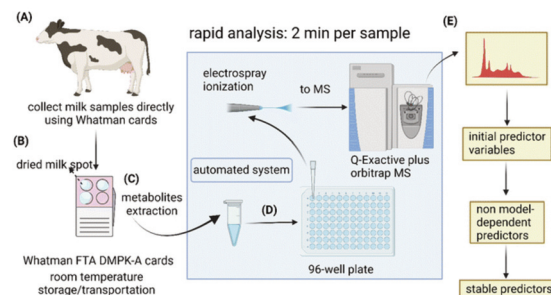
Typical applications

- Quantification of drug/drug metabolites, endogenous metabolites/lipids and contaminants in cells, microorganisms, plants, biofluids, tissues and environmental samples.
- Global metabolite profiling, metabolomics, biomarker discovery and surface analysis.
- Understanding the metabolic effects of biotic and/or abiotic perturbations on a biological system, for example drug-induced changes to intracellular metabolite pathways.
- Absolute quantification of a wide range of intra and extracellular metabolites using isotope dilution mass spectrometry.
- Isotope-assisted metabolic pathway profiling.

Metabolic alterations in dairy cattle with lameness revealed by untargeted metabolomics of dried milk spots using direct infusion-tandem mass spectrometry

To determine whether metabolic signatures associated with lameness could be discovered with untargeted metabolomics, we developed a novel workflow using direct infusion-tandem mass spectrometry to rapidly analyse (2 min per sample) dried milk spots (DMS) that were stored on commercially available Whatman® FTA® DMPK cards for a prolonged period (8 and 16 days). An orthogonal partial least squares-discriminant analysis (OPLS-DA) method validated by triangulation of multiple machine learning (ML) models and stability selection was employed to reliably identify important discriminative metabolites. With this approach, we were able to differentiate between lame and healthy cows based on a set of lipid molecules and several small metabolites.

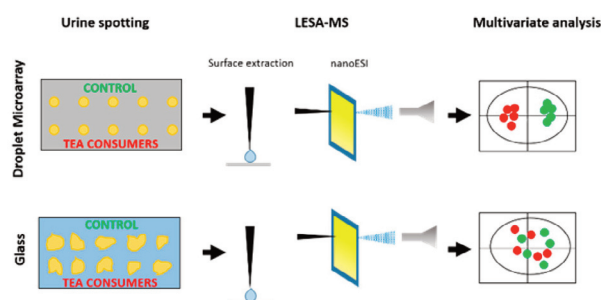
Wenshi He, Ana S. Cardoso, Robert M. Hyde, Martin J. Green, David J. Scurr, Rian L. Griffiths, Laura V. Randall and Dong-Hyun Kim, *Analyst*, 2022, 147, 5537



Size analysis of a mixture of polystyrene nanoparticles

The use of the Droplet Microarray (DMA) provides a surface-assisted LESA-MS method delivering significant improvement of the surface extraction repeatability leading to the acquisition of more robust and higher quality data. Such a method shows potential to be used for LESA-MS for controlled and reproducible surface extraction and for acquisition of high quality, qualitative data in a high-throughput manner.

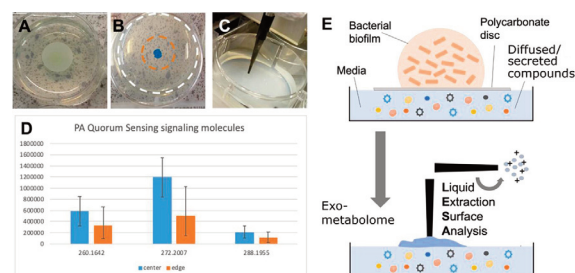
Meurs J, Alexander MR, Levkin PA, Widmaier S, Bunch J, Barrett DA and Kim D-H, *Anal. Chem.* 2018, 90, 10, 6001-6005



Probing interkingdom signalling molecules via Liquid Extraction Surface Analysis–Mass Spectrometry

Here, we present a model system for growing biofilms on discs before utilizing rapid and direct surface sampling MS, namely, liquid extraction surface analysis, to study the microbial exometabolome. One of the benefits of this approach is its surface-specific nature, enabling mimicking biofilm formation in a way that the study of planktonic liquid cultures cannot imitate. Our model system provides a route to investigate changes in the exometabolome, such as metabolites that become circulatory in the presence of multiple pathogens, and provides a rapid analytical approach to gaining a mechanistic understanding of bacterial signalling.

Shaun N. Robertson, Fadi Soukariéh, Thomas M. White, Miguel Camara, Manuel Romero, and Rian L. Griffiths, *Anal. Chem.* 2023, 95, 11, 5079–5086



Our facilities

QTRAP 6500+ and QTrap 4000 Quadrupole Linear Ion Trap LC-MS/MS

- Equipped with HPLC and UHPLC.
- Quantitative and qualitative targeted metabolite profiling, and for ID confirmation. Used where sensitivity is required.
- Advanced MS scanning options and information dependant acquisition offer unique approaches to metabolite profiling and identification.

Thermo Fisher QExactive (Hybrid quadrupole Orbitrap) high resolution mass spectrometer

- Equipped with NanoLC, UHPLC and TriVersa NanoMate.
- Small molecules profiling and identification.
- Quantitative and qualitative measurements.
- Metabolomics and lipidomics with high resolution measurements.

Thermo Orbitrap FusionTM LumosTM TribridTM Mass Spectrometer

- Resolution > 500,000 FWHM.
- MSn capabilities.

TriVersa NanoMate, Liquid Extraction Surface Analysis-Mass Spectrometry (LESA-MS)

- Surface extraction and analysis under ambient conditions.
- Direct infusion high throughput analysis.
- Advantages where UHV analysis may not be suitable; alternative to ToF-SIMS and MALDI.
- Coupled to QExactive MS to achieve a high resolution and confidence in identification using data dependant MS/MS.
- The new LESAPLUS allows for automated LESA experiments plus additional nano-LC separation.

Ambient Pressure Matrix Assisted Laser Desorption Ionisation (AP-MALDI)

- Surface analysis/imaging under ambient conditions.
- Unlike regular MALDI, it does not require a vacuum chamber, which means the analysis is faster and the samples can be kept in their native status.

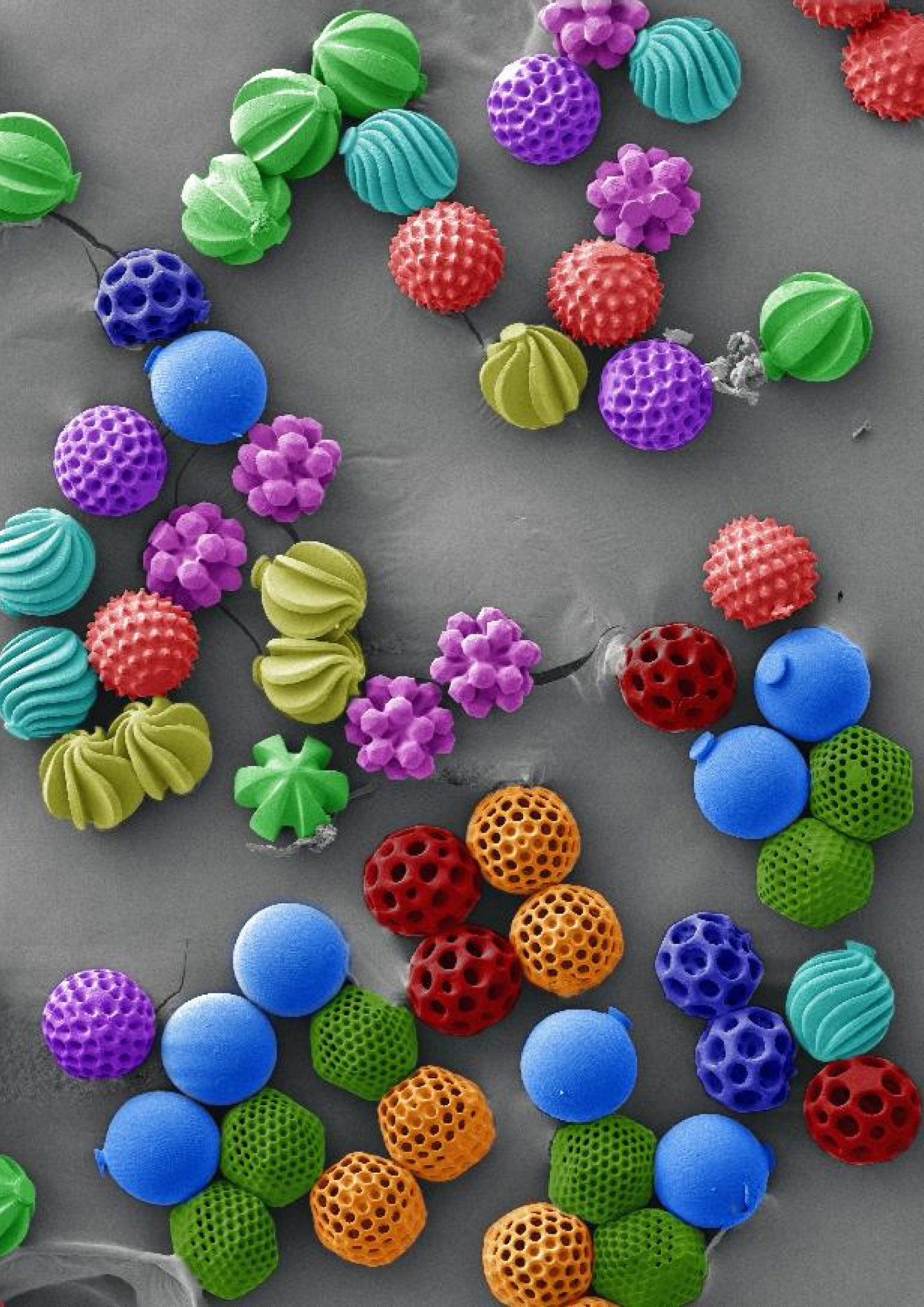
Data Analysis

- We perform data analysis for untargeted metabolomics with Compound Discoverer 3.3 (Thermo Scientific) and multivariate analysis with SIMCAP.
- For quantitative analysis we use TraceFinder (Thermo Fisher Scientific), Analyst (Sciex) and MultiQuant.

Find out how LCMS could help with your applications, designs or solutions:

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