

# Working Group 4 Update:

Development of Training Programs on Renal MRI

Andreas Pohlmann, Thoralf Niendorf







## WG4 Activities in 2019



Organization of ISMRM Member Initiated Symposium (MIS):

Frontiers in Magnetic Resonance Imaging Biomarkers of Renal Disease ISMRM Montreal 2019

Writing & editing of Springer Protocols book:

Preclinical MR Imaging of the Kidney: Methods and Protocols for Experiments and Analyses

Publication mid 2020



## Frontiers in Magnetic Resonance Imaging Biomarkers of Renal Disease

## **Organizers:**

- Thoralf Niendorf, Max-Delbrueck Center for Molecular Medicine, Berlin, Germany
- Octavia Bane, Icahn School of Medicine at Mount Sinai Hospital, New York, USA

Format: 7 presentations (each 14 min + 3 min discussion)

#### **Targets:**

- raise awareness for renal MRI
- outline opportunities, challenges, future directions of renal MRI
- · attract young scientists and new entrants into the field



- 1) The Link to Biology and Renal Physiology: The Physiologist's Perspective Erdmann Seeliger (M.D.), Charité University Medicine, Berlin, Germany
  - outline fundamentals of renal biology and renal physiology
- 2) Renal Diseases and Pathophysiology: The Nephrologist's Perspective Madhav Menon (M.D.), Icahn School of Med. at Mount Sinai Hospital, New York
  - presents unmeet clinical needs in nephrology



3) Emerging Renal MRI Biomarkers or Measurement Approaches: The MR Physics Perspective

Charlotte Buchanan (Ph.D.), Sir Peter Mansfield Imaging Centre, Nottingham, UK

- outlines current concepts and the physics of parametric MRI
- 4) Technical Validation: Demonstrating Accuracy, Precision and Quality Assurance of Renal MR Biomarkers

Ilona Dekkers (M.D.), Leiden University Medical Center, The Netherlands

highlights the challenges en route to MRI biomarkers of renal disease



- 5) Computational Models, Predictive Analytics and Machine Learning for Advancing Renal Diagnostics and Therapies
  - Satish Viswanath (Ph.D.), Case Western Reserve University, Cleveland, Ohio, USA
  - highlights the improvements in computational models and predictive analytics
- 6) Potential Added Value of Novel Renal MR Biomarkers in Drug Development or Patient Management
  - Lilach Lerman (M.D.), Mayo Clinic, Rochester, Minnesota, USA
  - discuss value of novel renal MRI biomarkers



## 7) Practical Challenges of Multi-center Studies and Clinical Renal MRI Trials

Paul Hockings (Ph.D.), Antaros Medical, Gothenburg, Sweden

#### MIS was great success and attracted large number of participants

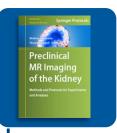
More than 200 attendees; lively discussions

## Motivated our plan for an ISMRM workshop on Renal MRI in April 2021

This was made part of the work program and deliverables of WG4

**Springer Protocols** Andreas Pohlmann Thoralf Niendorf Editors Preclinical **MR** Imaging ng lney of the Kidney **Methods and Protocols for Experiments** and Analyses

## **Book Project Addresses Two Current Limitations**



# **Lack of Training Material**

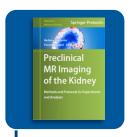
for students and new entrants to the field of renal MRI

## **Lack of Standardization**

Results from studies are difficult to compare

# **Objectives**





## **WG 4**

**Training** 

#### **WG 1**

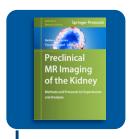
Standardization

# One-stop source for learning preclinical renal MRI

- Create library of protocols for a range of renal MR methods
- Protocols give detailed step-by-step instructions
- Complement experimental protocols with:
  - Advice on animal preparation & monitoring
  - Explanation of MRI method's basic concept
  - Instructions for data analysis

# **Objectives**





## **WG 4**

Training

#### **WG 1**

Standardization

## **Recommendations for preclinical renal MRI**

- First step towards harmonizing protocols
- MR parameter sets with rationales are convincing
- Supported by authors of several labs & PARENCHIMA
- Access to working protocol reduces need to "play-around"
- Paves the way for consensus-based recomm. in about 5yrs

# What are *Springer Protocols*?



- Methods in Molecular Biology (MIMB) is a critically acclaimed books series, part of the Springer Protocols
- Exists for 35 years: >2,100 books



with >47,000 protocols



- Each chapter (protocol) is provided in readily reproducible step-by-step fashion
  - 1. Short introduction
  - 2. List of **materials** needed
  - 3. Detailed **procedure**
  - 4. Comprehensive **notes** section (tips & tricks, troubleshooting advice)
- Each chapter is listed in **Pub** Med:

Assessment of Renal Hemodynamics and Oxygenation by Simultaneous Magnetic Resonance Imaging (MRI) and Quantitative Invasive Physiological Measurements.

Cantow K, Arakelyan K, Seeliger E, Niendorf T, Pohlmann A. Methods Mol Biol. 2016;1397:129-154. doi: 10.1007/978-1-4939-3353-2 11.

PMID: 26676132

## **Book Structure**



Book consists of 4 parts and 39 chapters (articles)

PART I Animal Models, Preparation, Monitoring and Physiological Interventions 3 chapters

PART II Measurement Techniques

12 chapters (Introductions the concepts and gives application examples)

PART III Experimental Protocols

13 sharters (Descriptions of experimental Protocols)

13 chapters (Descriptions of experimental steps)

**PART IV** Protocols for Advanced Analyses

11 chapters

# Unique features of this Springer Protocols book



- Experimental steps are described in generic terms
  - Rationale for choice of acquisition parameters is explained
  - Examples of specific parameter choices are given in *Notes* section
    - mouse & rat
    - animal & clinical scanner
    - different field strenghts

Each chapter/protocol is written by <u>authors from several labs</u>

# TOC

Address Springer Protocols

Address Street
Thank Street
Preclinical
MR Imaging
of the Kidney

Methods and Protocols for Experiments
and Analyses

Topics	PART I	PART II	PART III	PART IV
Animal Models of Renal Pathophysiology and Disease	Intro			
Preparation & Monitoring of Small Animals	Intro			
Reversible Physiological Interventions	Intro			
Invasive Probes for Quantitative Asst. of Renal Physiology		Concept	Exp. Prot.	
Multi-Modality Imaging (US, PET, Photoacoustic)		Concept		
MRI Hardware Considerations		Concept		
Essential Practical Steps (Slice planning, Shimming, TOF)			Exp. Prot.	
Renal Volume Measurement			Exp. Prot.	
T1 mapping		Concept	Exp. Prot.	Analysis
T2* & T2 mapping		Concept	Exp. Prot.	Analysis
DWI for ADC and IVIM		Concept	Exp. Prot.	Analysis
DCE-derived Perfusion and Filtration		Concept	Exp. Prot.	Analysis
ASL		Concept	Exp. Prot.	Analysis
CEST for Mapping of pH and Perfusion		Concept	Exp. Prot.	Analysis
<sup>23</sup> Na MRI		Concept	Exp. Prot.	Analysis
<sup>13</sup> C MR (Hyperpolarized)		Concept	Exp. Prot.	Analysis
<sup>19</sup> F Cell Tracking for Inflammatory Cell Migration		Concept	Exp. Prot.	Analysis
<sup>19</sup> F Oximetry			Exp. Prot.	
Subsegmentation of the Kidney (SOMBRERO, TLCO)				Analysis
Image Denoising for Parametric Mapping (NLM filter)				Analysis
	3	12	13	11

## **Preview**



## **Basic Concepts**

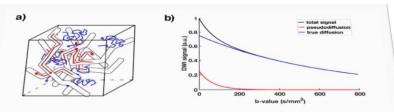


Figure 5: IVIM model. a) Schematic representation of random water motion in a voxel of renal tissue, where free diffusion component (in blue, described by the diffusion coefficient  $\mathcal{D}$ ) is complemented by fluid flowing in capillaries and tubules (in red, described by the pseudodiffusion coefficient  $\mathcal{D}^*$ ). b) Contributions of true diffusion and pseudodiffusion to the diffusion signal decay - pseudodiffusion is substantially faster than true diffusion, and so is only observed at low b-values.

## **Preview**



#### Rationales for parameters

a)

2. Set the shortest echo time (1E) and echo spacing (Δ1E) possible, under the condition that fat and water are in phase (see Note 2). The last TE should be close to the largest expected T<sub>2</sub>(\*\*) in the kidney multiplied by 1.5 (see Note 3). The aim is to acquire 10 or more echo images. Fewer TEs or larger ΔTE are not advisa kidney at high TEs can be so low that available for the analysis. Consider inc Fourier acceleration to shorten the first T

Figure 5: I renal tissu D) is comp pseudodi the diffus is only ob

- Choose the shortest possible repetition efficiency. TR will be limited by the le acquire.
- Adapt the flip angle (FA) to the TR and Ernst angle α<sub>E</sub> = arccos (exp(-TR/T<sub>1</sub>))
   larger FAs and determine the optimal I
- Set a high acquisition bandwidth (BV which decreases with the square root (see Note 5)
- Enable fat saturation. On ultrahigh fit the kidney due to chemical shift. At
- Enable the respiration trigger (per smotion blurring and unwanted inten TEs.

Parameter set examples

- 8. Example parameters for T<sub>2</sub>\* mapping of a 300g rat at 9.4T (Bruker small animal system): TR = 50 ms; flip angle = 16°; pulse length 1.0 ms; pulse bandwidth 5.4 kHz; receiver bandwidth = 109 kHz; number of echoes = 12; first echo = 2.14 ms; echo spacing 2.14 ms; TE = 2.14, 4.28, 6.42, 8.56, 10.7, 12.84, 14.98, 17.12, 19.26, 21.40, 23.54, 25.68 ms; averages = 4; slice orientation = coronal to kidney; frequency encoding = head-feet; FOV = (38.2x48.5) mm; matrix size = 169x115 zero-filled to 169x215; resolution = (0.226x0.421) mm; 1-3 slices with 1.4 mm thickness; fat suppression = on; respiration trigger = per slice; acquisition time = 40-60 s (with triggering under urethane anaesthesia).
- Example parameters for T<sub>2</sub>\* mapping of a 30g mouse at 4.7T (Agilent small animal system): TR = 350 ms; flip angle = 30°; receiver bandwidth = 100 kHz; number of echoes = 32; first echo = 2.0 ms; echo spacing 2.4 ms; TE = 2.00, 4.40, 6.80, 9.20, 11.60, 14.00, 16.40, 18.80, 21.20, 23.60, 26.00,..., 76.40 ms; averages = 4; slice orientation = coronal to kidney; frequency encoding = head-feet; FOV = (30x30) mm; matrix size = 128x128; resolution = (0.230x0.230) mm; 1 slice with 1.0 mm thickness; fat suppression = on; respiration trigger = on; acquisition time = 3.5-9.0 min (with triggering under isoflurane anaesthesia).
- 10. Example parameters for T<sub>2</sub>\* mapping of a 300g rat at 3.0T (Siemens Skyra<sup>fal</sup>, a clinical system): Animal position: Right decubitus; Coil: Knee; TR = 69 ms; flip angle = 30°; receiver bandwidth = 320 Hz/pixel; number of echoes = 12; first echo = 3.56 ms; echo spacing 3.43 ms; TE = 3.56, 6.99, 10.42, 13.85, 17.28, 20.71, 24.14, 27.57, 31.00, 34.43, 37.86, 41.29 ms;

## **Preview**



a)



Figure 5: I renal tissu D) is comp pseudodi the diffus is only ob b) 1

- 2. Set the shortest echo time (TE) and echo spacing (ΔTE) possible, under the condition the and water are in phase (see Note 2). The last TE should be close to the largest expected in the kidney multiplied by 1.5 (see Note 3). The aim is to acquire 10 or more echo in Fewer TEs or larger ΔTE are not advisa 8. Example parameters for Tx\* ways.
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total signal
 pseudodiffusion
 true diffusion

# 8. Example parameters for T<sub>2</sub> system): TR = 50 ms; flip angl bandwidth = 109 kHz; number = 2.14, 4.28, 6.42, 8.56, 10.7, 4; slice orientation = coronal mm; matrix size = 169x115 ze

with 1.4 mm thickness; fat sur

40-60 s (with triggering unde

- system): TR = 350 ms; flip and 32; first echo = 2.0 ms; echo 18.80, 21.20, 23.60, 26.00, frequency encoding = head (0.230x0.230) mm; 1 slice on; acquisition time = 3.5-9
- 10. Example parameters for system): Animal position: bandwidth = 320 Hz/pixel ms; TE = 3.56, 6.99, 10.42

Analysis with code examples

Example parameters for  $T_2^*$  mapping of a 200g ret at 0.4T (Poulon small animal system): TR = 50 ms; flip ang

- Define the model for fitting, where the two variables in X are SO and ADC (Note 14): curveADC = @(X,bVals) (X(1)\*exp(-bVals\*X(2)));
- Assign empty matrices for the two resulting maps, using the dimensions of the images (Note 15):
   SOmap = NaN (nRows, nCols);
   ADCmap = NaN (nRows, nCols);
- 3. For each voxel, loop through each row (I) and column (J) and extract the signal data
  for i = 1:nRows; for j = 1:nCols;
  Sb = img(i,j,:);
- 4. Estimate initial values for the variables to be fitted; use the b = 0 s/mm² value for SO, and approximate ADC by taking the gradient of the log signal between two points (Notes 16 & 17). See also section 3.2.4 for discussion of linearization of ADC estimation and fitting.
  SO\_est = Sb(1);
  ADC\_est = (log (max(Sb)) = log (min(Sb))) / (max(bVals) =

ADC\_est = (log(max(Sb))-log(min(Sb)))/(max(bVals)min(bVals));
X est = [S0 est ADC est];

 Run the least-squares curve fitting routine, using the initial estimates and providing lower and upper boundaries for the estimates if desired (Note 18). Assign the results to the allocated results maps:

```
limL = [0 0]; limU = [Inf Inf];
res = lsqcurvefit(curveADC, X_est, bVals, Sb, LimL, LimU);
SOmap(i,j) = res(1);
ADCmap(i,j) = res(2);
end; end; % close the row/column loops
```

## We are Close to the Finish Line!



Jan-Apr 2018

Finalizing concept  $\rightarrow$  proposal to *Springer*  $\rightarrow$  contract

May 2018

Assembling editorial team

Jul 2018

Editorial meeting via Skype

Jul-Aug 2018

Section editors identify & contact potential authors

Sept 2018

Disseminate chapter templates to authors

4 Oct 2018

Editors meeting @ Prague COST action meeting

4-5 Apr 2019

Apr - Sept 2019

Review process

Oct 2019

Formatting of chapters

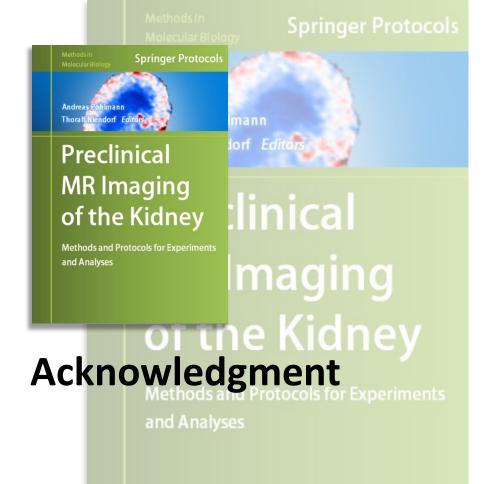
Nov 2019

Submission to Springer

Mid 2020

**Publication** 





## Turned into reality by ... 12 section editors



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## ... in 14 countries



Australia

Denmark

France

Germany

Hungary

Italy

Netherlands

Norway

Portugal

Spain

Sweden

Switzerland

UK

**USA** 



...to everyone who helped turning this vision into reality!